

Effects of Charcoal and Wood Vinegar Dietary Supplementation to Diarrhea Incidence and Faecal Hydrogen Sulfide Emissions in Pigs

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Abstract- Antibiotic resistance is now a big problem in livestock industry worldwide. Environmental pollution caused by the livestock industry has been becoming increasingly serious. Therefore, finding solutions to solve these problems simultaneously has become more necessary. Charcoal and wood vinegar has been used as fertilizer supplements and water filtration and treatment for humans and animals. This study aims to assess the applicability of charcoal and wood vinegar in prevention of diarrhea and reduction of environmental pollution in pig production. The experiments were conducted on pigs from weaned to finisher. The diets for pig were added different dosage of charcoal and wood vinegar (0.10; 0.20; 0.30% wood vinegar; 0.60; 0.80; 1.00% charcoal and 0.20 wood vinegar + 0.80 charcoal). The concentration of H₂S was measured by KITAGAWA-AP.20 kit. Results showed that charcoal and wood vinegar is effective in diarrhea prevention in pigs, reducing the percentage of pig's diarrhea. Moreover, the concentrate of H₂S in pig houses decreased in the experiment groups of adding charcoal and wood vinegar (0.30% wood vinegar, 0.60% and 0.80% charcoal, and 0.20% wood vinegar+0.80% charcoal) to diets. Therefore, the study results confirm the important roles of both charcoal and wood vinegar used as additives to diet for pigs.

Index Terms- charcoal, diarrhea, wood vinegar

I. INTRODUCTION

Diarrhea is widespread disease causes great economic loss in the swine industry worldwide [2, 19]. Diarrhea can occur at any developmental stages of pigs, from suckling piglet to slaughter pigs. Diarrhea can be caused by nutrition, bacteria, viruses, parasites or combination of those agents. This disease is responsible for economic losses due to high mortality, morbidity, and cost of medicine application as well as decreasing growth performance in culturing pig. Previously, *Salmonella* spp., *Clostridium perfringens*, *Serpulina* *hyodysenteriae* have been known as diarrhea-related pathogens [7]. *Escherichia coli* is evidenced as the main pathogen causing diarrhea in cultured pig. The factors that contribute to the increasing of number of diarrhea outbreaks associated with *E. coli* are not yet fully understood [10].

Antibiotics had widely used in animal culture industry, throughout the contributing the great success of animal

agriculture after World War II. However, the application of antibiotics in livestock has stimulated the resistant ability with antibiotics in bacterial pathogens [1, 17, 33]. To control this scenario supplementation of exogenous additives, including egg yolk from immunized chickens, zinc and/or spray-dried plasma or probiotics to diets as well as dietary acidification and phage therapy have been intruded and previously applied. To date, a single control strategy has unsuccessful to prove the total effectiveness; a combination of diet change and other preventive measures are necessary to apply [10].

Hydrogen sulfide (H₂S) emission from pig culture system associates with several problems, including biogenic corrosion of concrete, release of obnoxious odors to the urban atmosphere and toxicity of sulfide gas to farmers [23, 44]. Environmental pollution caused by livestock production has been much research to show. Animal waste containing some compounds have been confirmed causing environmental pollution [29, 43], especially, concerning the pollution caused by nitrogen in feces of pig culture industry [24]. Due to their physical attributes where pigs only consume about 50% components of nitrogen, phosphorus and sulfur in the diets [28, 34], decreasing H₂S emissions from pig production is a globally considered matter. The chemical composition of livestock waste, volatile gases emitted into the environment depends on the control of diets [19]. The impact on crude protein, amino acids in the diet are able to restrict odor intensity and emissions from pig manure [12, 15, 32]. The sulfur-containing compounds are the most important agents caused stench from manure. Changing diets in culturing system through controlling the crude protein content, composition and proportion of fiber can reduce the emission of H₂S from manure [31]. According to Bindelle, *et al.* [4], the changes of the fiber content in the diet for pigs could reduce NH₃ elimination of shelters by reducing the amount of nitrogen in the urine.

Wood vinegar also called pyrolygneous acid or liquid smoke is the water condensate of smoke produced during wood carbonization. To completely exploit the sawdust, branch and shell of the forestry industry, most of these wood materials are used to produce charcoal [5]. Wood vinegar is a complex mixture of water, phenols, guaiacols, vanillins, catecols, syringols, furan carboxaldehydes, isoeugenol, pyrones, acetic acid, formic acid and other carboxylic acids [20]. Charcoal had been known as a good adsorption material because of easy to

linking to different types of molecules [6]. Charcoal has been previously proved to be very efficient in removal of bacteria and bacterial toxins in both *invivo* and *in vitro* studies [8-9, 11, 26]. Naka, *et al.*, [21] demonstrated that charcoal could remove the toxins of *E. coli*. Other authors used wood vinegar to inhibit the growth of bacteria involving in intestinal disease odd [3, 13, 22], as well as to control the presence of *Cryptosporidium parvum* in the digestive tract of ruminants [14]. Yanyong and Sukhumwat, [40] concluded that wood vinegar is effective in inhibiting the grow and pathogenicity of pathogens (fungi and bacteria) on plants.

II. MATERIALS AND METHODS

Animals and diets

A total of 32 weaned breed F1 pigs (Landrace x MongCai) were allocated to 8 groups. Each of one pig was housed in a cage of 0.8x0.6x0.5m equipped automatic drinking valves. The similarity of body weight of pigs between treatments was ensured. The initial weight of pigs in each treatment is presented

in Table 1. All pigs were vaccinated before conducting experiment.

Charcoal and wood vinegar was produced as described below. Wood vinegar liquid obtained after cooling charcoal smoke from wood of *Acacia auriculiformis* trees by dry distillation at 350 to 450°C was kept for 1 to 2 years. Then the filtrate solution was distilled to remove harmful substances such as tar. The experimental diets were formulated to meet all nutrition requirements for pigs from weaned to 30kg-weight period and from 30kg-weight period to finisher according to PHILSAN's recommendation (2003). The ingredients in the diets include rice bran, maize, fish meal, cassava meal, rice bran and mineral-vitamin premix. A total of 7 test diets were established based on control diets by adding 3 dosages of wood vinegar (0.10%, 0.20% and 0.30%), 3 dosages of charcoal (0.60%, 0.80% and 10%), and mixture of 0.20% wood vinegar and 0.80% of charcoal. The experiments were conducted in three replicate. The composition and nutrient content of experimental diets are presented in Tables 2 and 3.

Table 1: The initial body weight of pigs

Treatment	Weight of pigs	Number of pigs per cage	Number of repeats
Control	8.87 ± 0.61	1	4
0.1% wood vinegar	9.35 ± 0.73	1	4
0.2% wood vinegar	8.65 ± 0.74	1	4
0.3% wood vinegar	8.48 ± 0.35	1	4
0.6% charcoal	9.15 ± 0.65	1	4
0.8% charcoal	8.65 ± 0.18	1	4
1.0% charcoal	9.20 ± 0.27	1	4
0.2% wood vinegar + 0.8% charcoal	9.18 ± 0.66	1	4

Experimental procedure

Pigs were fed *ad libitum* twice a day (7:00 and 17:00) and had free access to water during experiment. Data of pigs with diarrhea had also daily recorded in the morning. The diarrhea cases recorded based on the symptoms was described by Will [39]. The data of diarrhea of pig was recorded from start to finish experiment. Before concentration H₂S was measuring began, a small test was done to determine the sample weight and timing of suitable with kit. Over a period of 5 days (from 45th to 49th day of experiment), faeces from each pig in both phases (weaning to

30kg-weight period, and 30kg-weight to finisher period) were collected immediately after spontaneous defecation. Faeces of 4 pigs within a replication were pooled. Then 200 grams of faecal samples was subsampled and placed in two 7.5 cm diameter x 15cm height plastic bottle for measuring of H₂S. All the bottles had been predrilled a hole of 0.5mm diameter on the cap to create aerobic conditions. Then, concentration of H₂S was measured using KITAGAWA-AP.20 kit (Kitagawa, Japan).

Table 2. Composition and nutrient content of experimental diets for pigs from weaned to 30kg

Ingredients	Treatments							
	Control	0.1% wood vinegar	0.2% wood vinegar	0.3% wood vinegar	0.6% charcoal	0.8% charcoal	1.0% charcoal	0.2% wood vinegar + 0.8% charcoal
Ingredients								
Maize (%)	29.55	29.55	29.55	29.55	29.55	29.55	29.55	29.55
Cassava meal (%)	14.62	14.62	14.62	14.62	14.62	14.62	14.62	14.62
Fish meal (%)	17.91	17.91	17.91	17.91	17.91	17.91	17.91	17.91
Rice bran (%)	35.70	35.70	35.70	35.70	35.70	35.70	35.70	35.70

Mineral –vitamin premix (%)	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50
Charcoal (%)	0.00	0.00	0.00	0.00	0.60	0.80	1.00	0.80
Wood vinegar (%)	0.00	0.10	0.20	0.30	0.00	0.00	0.00	0.20
Chemical composition								
Crude protein (%)	18.00	18.00	18.00	18.00	18.00	18.00	18.00	18.00
ME (kcal/kg)	3265	3265	3265	3265	3265	3265	3265	3265
Lysin (%)	1.01	1.01	1.01	1.01	1.01	1.01	1.01	1.01
Met + Cys (%)	0.58	0.58	0.58	0.58	0.58	0.58	0.58	0.58

One hundred grams of mineral – vitamin premix contains 10,800 Mn, 2,160 mg Fe, 7,200 mg Zn, 1,260 mg cu, 144 mg Iodine, 21.6 mg Co, 14.1 mg Se, 40 mg acid folic, 4,800 mcg biotin, 20,000 mg choline chloride

Table 3. Composition and nutrient content of experimental diets for pigs from 30kg to slaughter

Ingredients	Treatments							
	Control	0.1% wood vinegar	0.2% wood vinegar	0.3% wood vinegar	0.6% charcoal	0.8% charcoal	1.0% charcoal	0.2% wood vinegar + 0.8% charcoal
Ingredients								
Maize (%)	21.01	21.01	21.01	21.01	21.01	21.01	21.01	21.01
Cassava meal (%)	17.33	17.33	17.33	17.33	17.33	17.33	17.33	17.33
Fish meal (%)	10.82	10.82	10.82	10.82	10.82	10.82	10.82	10.82
Rice bran (%)	50.36	50.36	50.36	50.36	50.36	50.36	50.36	50.36
Mineral –vitamin premix (%)	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50
Charcoal (%)	0.00	0.00	0.00	0.00	0.60	0.80	1.00	0.80
Wood vinegar (%)	0.00	0.10	0.20	0.30	0.00	0.00	0.00	0.20
Chemical composition								
Crude protein (%)	15.50	15.50	15.50	15.50	15.50	15.50	15.50	15.50
ME (kcal/kg)	3265	3265	3265	3265	3265	3265	3265	3265
Lysin (%)	0.66	0.66	0.66	0.66	0.66	0.66	0.66	0.66
Met + Cys (%)	0.39	0.39	0.39	0.39	0.39	0.39	0.39	0.39

One hundred grams of mineral – vitamin premix contains 10,800 Mn, 2,160 mg Fe, 7,200 mg Zn, 1,260 mg cu, 144 mg Iodine, 21.6 mg Co, 14.1 mg Se, 40 mg acid folic, 4,800 mcg biotin, 20,000 mg choline chloride.

Statistical analysis

The data obtained from experiment were analyzed using general linear model (GLM) in SPSS software version 18.0. Significant differences among means were separated by Tukey test or χ^2 with a 5% level of probability. The frequency of diarrhea

was expressed as percentage of days with diarrhea for each pig. The frequency of diarrhea of pig was calculated by the following formula. (%day/pig=(total days with diarrheaof pig/total days of the experiment)*100).

III. RESULTS AND DISCUSSION

Growth performance

Table 4. Growth performance of pigs in the experiment

	Initial BW (kg)	Finish (before 30kg phase) BW (kg)	Final BW (kg)	ADG from weaned to 30kg (g/day)	ADG from 30kg to slaughter (g/day)	ADG from weaned to slaughter (g/day)
Control	8.87	37.25	71.50	378.33 ^a	456.67 ^a	417.50 ^a
0.10% WG	9.35	42.93	82.93	447.78 ^b	533.33 ^a	490.56 ^b
0.20% WG	8.65	40.40	79.15	423.33 ^{ab}	516.67 ^a	470.00 ^{bc}
0.30% WG	8.48	39.98	77.86	420.00 ^{ab}	505.00 ^a	462.50 ^{bc}

0.60% C	9.15	37.82	78.48	382.22 ^a	495.17 ^a	438.69 ^{ac}
0.80% C	8.65	37.90	73.03	390.00 ^{ab}	468.33 ^a	429.17 ^{ac}
1.00% C	9.20	37.33	73.08	375.00 ^a	476.67 ^a	425.83 ^a
0.20% WG+0.80% C	9.18	38.93	75.18	396.67 ^{ab}	483.33 ^a	440.00 ^{ac}

^{a-c}Means with different superscripts within a row differ significantly ($p < 0.05$).

CT: Control; WG: Wood vinegar; C: Charcoal; BW: body weight; ADG: Average daily gain

The growth performance results show that, the daily gain of 0.10% wood vinegar group was highest (490.56g/day), control group was lowest (417.50g/day). The daily gain of 0.10; 0.20 and 0.30% wood vinegar group were higher control group and 1.00% charcoal group ($P < 0.05$). With increasing dietary wood vinegar

levels, daily gain tended to decrease, but no significant ($P > 0.05$). Meanwhile, daily gain of dietary charcoal levels lower dietary wood vinegar levels and mix charcoal and wood vinegar.

Effects of charcoal and wood vinegar to diarrhea incidence in pigs

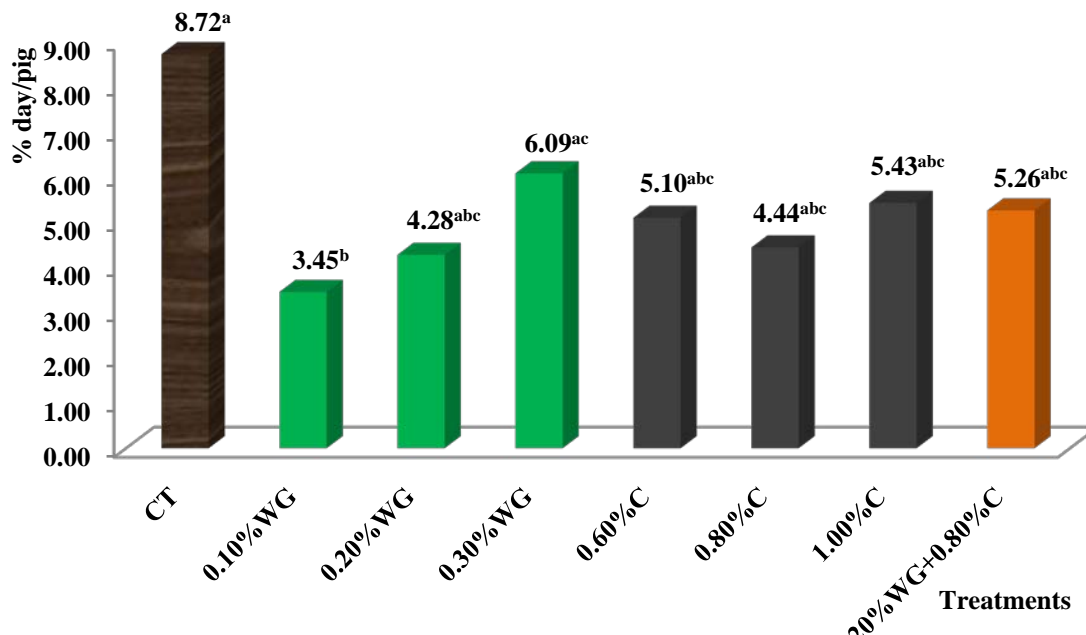


Figure 1: Frequency of diarrhea (% day/pig)

^{a-c} Means with different superscripts within a column differ significantly ($P < 0.05$).

CT: Control; WG: Wood vinegar; C: Charcoal

The results in Fig. 1 show the frequency of diarrhea in pigs during experiment period. The highest frequency of diarrhea was observed in the control group (8.72% day/pig). Meanwhile, the lowest percentage days (3.45%) with diarrhea of pigs was found in the group of fed on the diet added 0.10% wood vinegar. The difference in frequency of diarrhea in pigs fed on control diet and diet added 0.10% wood vinegar was significant ($P < 0.05$). There was no significant difference in frequency of diarrhea in pigs between the 6 remain treatments (Fig. 1). The increasing tendency was observed for the frequency of diarrhea in pigs fed on wood vinegar with increasing concentrations. The percentage of 3.45, 4.28 and 6.09% day of pigs got diarrhea was observed when fed on diets added 0.10, 0.20 and 0.30% wood vinegar, respectively. Compared to pigs fed control diet, the pigs fed on diets supplemented 0.20% wood vinegar and 0.80% charcoal showed the decrease of 39.7% in occurrences of diarrhea. In the treatments use charcoal as an additive to the diets of experimental pigs, the percentage (% day/pig) of diarrhea odd

was lower than the controls; in the treatment of 0.6%, 0.8% and 1.00% charcoal was 5.10, 4.44 and 5.43, but in the control group was 8.72 (% day/pig). This study results are consistent to previous study results.

The usefulness of activated charcoal was reported in reducing the effects of toxins in diets by preventing their absorption through the intestine [25-27, 37]. Watarai and Tana, [36] indicated the mixture of wood vinegar and charcoal reduce the virulence and numbers of *Salmonella* and increase the number of useful bacteria (*E. faecium* and *B. thermophilum*) in the digestive tract of poultry. Wood vinegar compound liquid was induced acetic acid, propanoic acid, butanoic acid, dimethylphenol and methoxyphenol [42][30]. Which organic acids included in wood vinegar solutions controlled the balance of intestinal microflora and pathogens [18, 27, 36]. Yanyong and Sukhumwat, [40] evaluated the effect of adding a mixture of charcoal and wood vinegar to feed to the gastrointestinal mucosa of chickens and resulted in at the 1% mixture of food, less

change mucosal surfaces; but at the 5%, microvilli eroded. [Naka et al., \[21\]](#) recommended charcoal could be used to remove the toxins excreted from *E. coli* O157:H7 after 5 minutes mixing 10 mg charcoal into the liquid of bacterial-growing media. The protective efficacy of charcoal and wood vinegar against intestinal infection with some of bacterial, fungal and parasites was proven [5, 8-9, 16, 21, 36], some of which are causes of diarrhea in pig. Furthermore, [Paraud et al., \[25\]](#) found that charcoal was able to use for preventing disease caused by *C. parvum*

goats. Combined these above results, it is obviously that charcoal and wood vinegar added to diets for pigs are good and validable to prevent porcine diarrhea. Charcoal could well adsorb toxins and other components excreted by pathogen bacteria; wood vinegar could improve the digestibility and nutritional absorption, inhibit the growing of pathogenic bacteria and stimulate the growing of potentially useful bacteria harbored in the gastrointestinal tract of pigs.

H₂S emissions from pig's manure

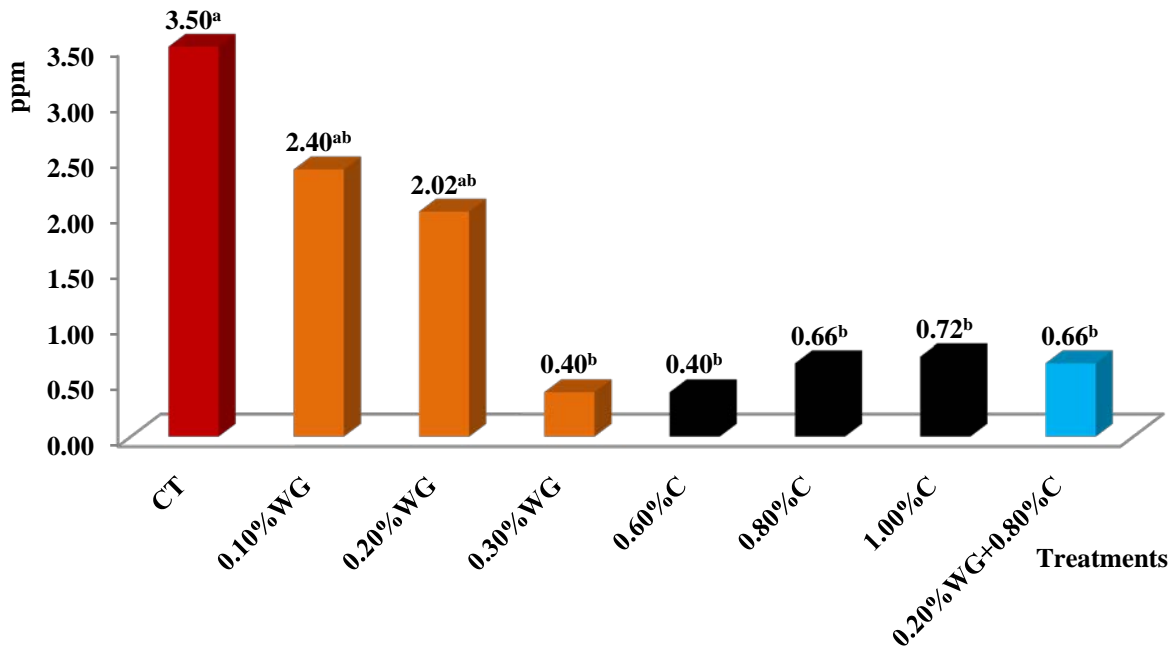


Figure 2: The concentration of H₂S emissions from manure of pig from weaned to 30kg

^{a-c} Means with different superscripts within a column differ significantly ($P < 0.05$).

CT: Control; WG: Wood vinegar; C: Charcoal

Concentrations of H₂S were measured after 4 hours post sampling. The results showed that the highest concentration of H₂S was found in the control group 3.50ppm, and at 0.10% and 0.20% vinegar of the treatments 2.40 and 2.02ppm respectively ([Fig. 2](#)). In contrast, the lowest concentration of H₂S was determined in the treatments of 0.30% vinegar and 0.60% charcoal, sharing a similar concentration of 0.40ppm. Statistically, the average concentration of H₂S in treatments, excepting for 0.10% and 0.20% vinegar treatments, was significantly different ($P < 0.05$) in comparison with the control groups. The results also show that an opposite relation between the doses of vinegar to the concentrations of H₂S measured in faeces of pigs. However, it is found to be different in the treatment of supplementation of charcoal, the increase of dose uses, the high concentrations of H₂S. Additionally, in the treatment of 0.20% vinegar + 0.80% charcoal, the concentration

of H₂S was lower than the controls, it also lower than treatments 0.20% vinegar, but higher treatments 0.80% charcoal.

The results of H₂S concentration emitted in the second phase of the experiment ([from](#) 30kg-weight to ending) show that it was the highest (2.82ppm) in the control group and the lowest (0.98ppm) in the treatment of 0.80% charcoal ([Fig. 3](#)). The concentrations of H₂S tended to decrease with the increasing percent of vinegar added to the diets. The concentrations of H₂S in all treatments were lower than the control group, but not statistically significant ($P < 0.05$). In the treatment of 0.20% vinegar + 0.80% charcoal, the average concentration of H₂S (1.40ppm) was lower than the control (2.82ppm). Moreover, the average concentration of H₂S in the treatments of 0.20% vinegar (1.96ppm) was higher than such in the treatments 0.80% charcoal (0.98ppm). The results suggest the existence of interaction between the use of charcoal and wood vinegar when added to diets to decrease the H₂S emissions from manure pigs.

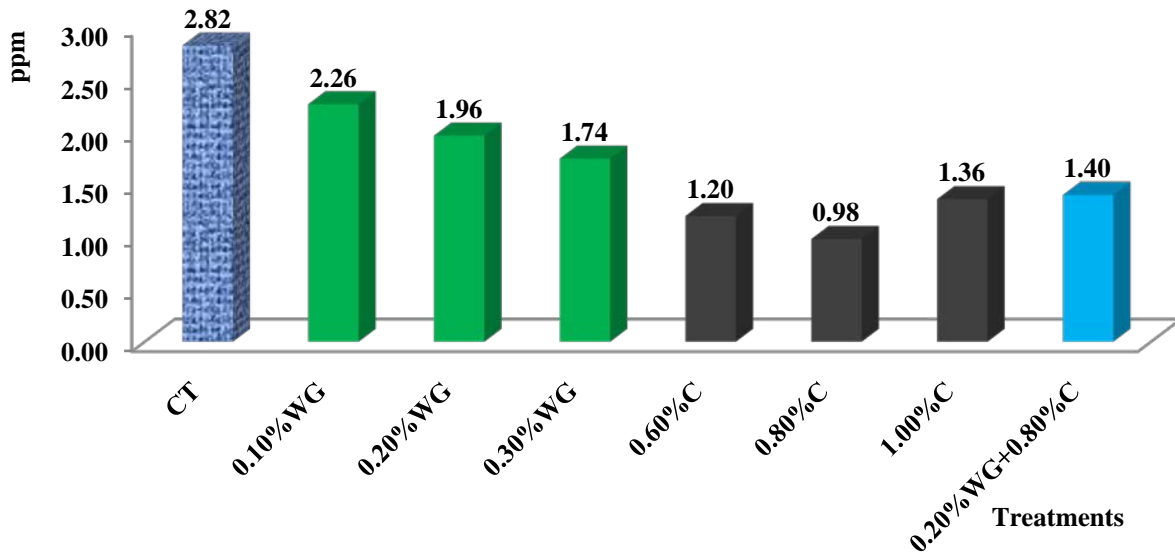


Figure 3: The concentration of H₂S emissions from manure of pig from 30kg to finisher

^{a-c} Means with different superscripts within a column differ significantly (p < 0.05).

CT: Control; WG: Wood vinegar; C: Charcoal

[Whitehead, et al.](#) has reported that quebracho tannins added to diets of swine reduced overall gas, hydrogen sulfide, and methane production by greater than 90% after 7 days of treatment and continued to at least 28 days post treatment as well as decreased the total bacterial population in the gastrointestinal tract. In our study, charcoal added to pig diets could reduce the concentration of H₂S from 2-8.5 times, being accordant to previous study results that a removing of 98% H₂S gas produced by pig manure using bamboo charcoal as an additive to diets [41]. Additionally, [Wang, et al.](#), were conducted to evaluate the effects of the dietary with BioPlus 2B® supplemented on the concentration of H₂S emitted from growing pigs; however, in this study, the H₂S emissions were not affected by supplementation with BioPlus 2B®.

IV. CONCLUSION

This study provides basic information on the advantages of application of charcoal and wood vinegar as additive components in diets of pig during life stages. The current study confirmed these additives functioning in the prevention of porcine diarrhea as well as a reduction of H₂S emissions excreted from pig products. However, there should be more experiment to clarify the mechanism of action as well as expand the applications of preparations towards this. In this study only evaluated some doses of charcoal and wood vinegar for purposes of prevention of diarrhea and reduce emissions of H₂S. So, research on a large scale and other dose of charcoal and wood vinegar addition to the diets need to further perform.

REFERENCES

[1] Aalbaek, B., Rasmussen, J., Nielsen, B., Olsen, J.E. (1991). Prevalence of antibiotic-resistant *Escherichia coli* in Danish pigs

and cattle. *APMIS : acta pathologica, microbiologica, et immunologica Scandinavica*. 99(12): 1103-1110.

[2] Alvarez, J., Sarradell, J., Morrison, R., Perez, A. (2015). Impact of porcine epidemic diarrhea on performance of growing pigs. *PLoS one*. 10(3): 1-8.

[3] Anderson, M.E., Marshall, R.T., Dickson, J.S. (1991). Efficacies of acetic, lactic and two mixed acids in reducing number of bacteria on surface of lean meat. *Journal of Food Safety*. 12(2): 139-147.

[4] Bindelle, J., Buldgen, A., Delacollette, M., Wavreille, J., Agneessens, R., Destain, J., P, Leterme, P. (2009). Influence of source and concentrations of dietary fiber on in vivo nitrogen excretion pathways in pigs as reflected by in vitro fermentation and nitrogen incorporation by fecal bacteria. *Journal of animal science*. 87(2): 583-593.

[5] Cai, K., Jiang, S., Ren, C., He, Y. (2012). Significant damage-rescuing effects of wood vinegar extract in living *Caenorhabditis elegans* under oxidative stress. *J Sci Food Agric*. 92(1): 29-36.

[6] Chandy, T., Sharma, C.P. (1998). Activated charcoal microcapsules and their applications. *Journal of biomaterials applications*. 13(2): 128-157.

[7] Chiu, C.H., Su, L.H., Chu, C. (2004). *Salmonella enterica* Serotype Choleraesuis: Epidemiology, Pathogenesis, Clinical Disease, and Treatment. *Clinical Microbiology Reviews*. 17(2): 311-322.

[8] Drucker, M.M., Goldhar, J., Ogra, P.L., Neter, E. (1977). The effect of attapulgite and charcoal on enterotoxicity of *Vibrio cholerae* and *Escherichia coli* enterotoxins in rabbits. *Infection*. 5(4): 211-213.

[9] Du, X.N., Niu, Z., Zhou, G.Z., Li, Z.M. (1987). Effect of activated charcoal on endotoxin adsorption. Part I. An in vitro study. *Biomaterials, artificial cells, and artificial organs*. 15(1): 229-235.

[10] Fairbrother, J.M., Nadeau, E., Gyles, C.L. (2005). *Escherichia coli* in postweaning diarrhea in pigs: an update on bacterial types, pathogenesis, and prevention strategies. *Animal health research reviews / Conference of Research Workers in Animal Diseases*. 6(1): 17-39.

[11] Gardiner, K.R., Anderson, N.H., McCaigue, M.D., Erwin, P.J., Halliday, M.I., Rowlands, B.J. (1993). Adsorbents as antiendotoxin agents in experimental colitis. *Gut*. 34(1): 51-55.

[12] Gralapp, A.K., Powers, W.J., Faust, M.A., Bundy, D.S. (2002). Effects of dietary ingredients on manure characteristics and odorous emissions from swine. *Journal of animal science*. 80(6): 1512-1519.

[13] Hsiao, C.P., Siebert, K.J. (1999). Modeling the inhibitory effects of organic acids on bacteria. *International Journal of Food Microbiology*. 47(3): 189-201.

[14] Kniel, K.E., Sumner, S.S., Lindsay, D.S., Hackney, C.R., Pierson, M.D., Zajac, A.M., Golden, D.A., Fayer, R. (2003). Effect of organic acids and

- hydrogen peroxide on *Cryptosporidium parvum* viability in fruit juices. Journal of food protection. 66(9): 1650-1657.
- [15] Li, Q.F., Trottier, N., Powers, W. (2015). Feeding reduced crude protein diets with crystalline amino acids supplementation reduce air gas emissions from housing. Journal of animal science. 93(2): 721-730.
- [16] Li, X.G., Han, L.H., Wu, S.Q., Piao, R.Z., Liu, H.F. (2014). Inhibition effects of wood vinegar on *Alternaria panax* and *Botrytis cinerea*. Zhong Yao Cai. 37(9): 1525-1528.
- [17] Mathew, A.G., Upchurch, W.G., Chattin, S.E. (1998). Incidence of antibiotic resistance in fecal *Escherichia coli* isolated from commercial swine farms. Journal of animal science. 76(2): 429-434.
- [18] Mekbungwan, A., Yamauchi, K., Sakaida, T., Buwjoom, T. (2008). Effects of a charcoal powder-wood vinegar compound solution in piglets for raw pigeon pea seed meal. Animal. 2(3): 366-374.
- [19] Miller, D.N., Varel, V.H. (2003). Swine manure composition affects the biochemical origins, composition, and accumulation of odorous compounds. Journal of animal science. 81(9): 2131-2138.
- [20] Mohan, D., Pittman, C.U., Steele, P.H. (2006). Pyrolysis of Wood/Biomass for Bio-oil: A Critical Review. Energy & Fuels. 20(3): 848-889.
- [21] Naka, K., Watarai, S., Tana, Inoue, K., Kodama, Y., Oguma, K., Yasuda, T., Kodama, H. (2001). Adsorption effect of activated charcoal on enterohemorrhagic *Escherichia coli*. The Journal of veterinary medical science / the Japanese Society of Veterinary Science. 63(3): 281-285.
- [22] Nakai, S.A., Siebert, K.J. (2003). Validation of bacterial growth inhibition models based on molecular properties of organic acids. Int J Food Microbiol. 86(3): 249-255.
- [23] Nielsen, P.H., Raunkjaer, K., Hvitved Jacobsen (1998). Sulfide production and wastewater quality in pressure mains. Water Science Technology. 37(97-10).
- [24] Otto, E.R., Yokoyama, M., Ku, P.K., Ames, N.K., Trottier, N.L. (2003). Nitrogen balance and ileal amino acid digestibility in growing pigs fed diets reduced in protein concentration. Journal of animal science. 81(7): 1743-1753.
- [25] Paraud, C., Pors, I., Journal, J.P., Besnier, P., Reisdorffer, L., Chartier, C. (2011). Control of cryptosporidiosis in neonatal goat kids: efficacy of a product containing activated charcoal and wood vinegar liquid (Obioneck(R)) in field conditions. Veterinary parasitology. 180(3-4): 354-357.
- [26] Pegues, A.S., Sofer, S.S., McCallum, R.E., Hinshaw, L.B. (1979). The removal of ¹⁴C labeled endotoxin by activated charcoal. The International journal of artificial organs. 2(3): 153-158.
- [27] Samanya, M., Yamauchi, K. (2001). Morphological changes of the intestinal villi in chickens fed the dietary charcoal powder including wood vinegar compounds. Journal of Poultry Science. 4(38): 289-301.
- [28] Sands, J.S., Ragland, D., Baxter, C., Joern, B.C., Sauber, T.E., Adeola, O. (2001). Phosphorus bioavailability, growth performance, and nutrient balance in pigs fed high available phosphorus corn and phytase. Journal of animal science. 79(8): 2134-2142.
- [29] Schiffman, S.S., Bennett, J.L., Raymer, J.H. (2001). Quantification of odors and odorants from swine operations in North Carolina. Agricultural and Forest Meteorology. 108(3): 213-240.
- [30] Shi Qi Zeng, Lei Jun Feng, Hui, W.Z. (2008). Analysis on organic components of wood vinegar of applewood. Chinese Journal of Analysis Laboratory. 27(10): 70-72 in Chinese.
- [31] Sutton, A.L., Kephart, K.B., Verstegen, M.W., Canh, T.T., Hobbs, P.J. (1999). Potential for reduction of odorous compounds in swine manure through diet modification. Journal of animal science. 77(2): 430-439.
- [32] Trabue, S., Kerr, B. (2014). Emissions of greenhouse gases, ammonia, and hydrogen sulfide from pigs fed standard diets and diets supplemented with dried distillers grains with solubles. Journal of environmental quality. 43(4): 1176-1186.
- [33] van Den Bogaard, A.E., London, N., Stobberingh, E.E. (2000). Antimicrobial resistance in pig faecal samples from the Netherlands (five abattoirs) and Sweden. The Journal of antimicrobial chemotherapy. 45(5): 663-671.
- [34] van Kempen, T.A., Baker, D.H., van Heugten, E. (2003). Nitrogen losses in metabolism trials. Journal of animal science. 81(10): 2649-2650.
- [35] Wang, Y., Cho, J.H., Chen, Y.J., Yoo, J.S., Huang, Y., Kim, H.J., Kim, I.H. (2009). The effect of probiotic BioPlus 2B® on growth performance, dry matter and nitrogen digestibility and slurry noxious gas emission in growing pigs. Livestock Science. 120(1-2): 35-42.
- [36] Watarai, S., Tana (2005). Eliminating the carriage of *Salmonella enterica* serovar Enteritidis in domestic fowls by feeding activated charcoal from bark containing wood vinegar liquid (Nekka-Rich). Poultry science. 84(4): 515-521.
- [37] Watarai, S., Tana, Koiwa, M. (2008). Feeding activated charcoal from bark containing wood vinegar liquid (nekka-rich) is effective as treatment for cryptosporidiosis in calves. Journal of dairy science. 91(4): 1458-1463.
- [38] Whitehead, T.R., Spence, C., Cotta, M.A. (2013). Inhibition of hydrogen sulfide, methane, and total gas production and sulfate-reducing bacteria in in vitro swine manure by tannins, with focus on condensed quebracho tannins. Applied microbiology and biotechnology. 97(18): 8403-8409.
- [39] Wills, R.W. (2000). Diarrhea in growing-finishing swine. Vet Clin North Am Food Anim Pract. 16(1): 135-161.
- [40] Yanyong, C., Sukhumwat, P. (2009). Wood vinegar: by product from rural charcoal kiln and its role in plant protection. Asian Journal of Food and Agro-Industry. S189-S195(Special Issue): 189-195.
- [41] Ying, C.C., Yu, Y.L., Ching, P.T. (2004). Control of H₂S waste gas emissions with a biological activated carbon filter. Journal of Chemical Technology and Biotechnology. 79(6): 570-577.
- [42] Yoshimura, H., Washio, H., Yoshida, S., Seino, T., Otaka, M., Matsubara, K., Matsubara, M. (1995). Promoting effect of wood vinegar compounds on fruit-body formation of *Pleurotus ostreatus*. Mycoscience. 36(2): 173-177.
- [43] Zahn, J.A., DiSpirito, A.A., Do, Y.S., Brooks, B.E., Cooper, E.E., Hatfield, J.L. (2001). Correlation of human olfactory responses to airborne concentrations of malodorous volatile organic compounds emitted from swine effluent. Journal of environmental quality. 30(2): 624-634.
- [44] Zhang, L., De Schryver, P., De Gussem, B., De Muynck, W., Boon, N., Verstraete, W. (2008). Chemical and biological technologies for hydrogen sulfide emission control in sewer systems: a review. Water research. 42(1-2): 1-12.

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