

Medicinal uses and Pharmacological activities of *Cyperus rotundus* Linn – A Review

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Abstract- *Cyperus rotundus* Linn belong to the family Cyperaceae. It is the world worst weed native to India. It grows in small clump up to 100cm high. The extensive distribution of the nut-grass is due to its ability to adapt to a wide range of soil types, altitudes, temperatures, soil pH and moisture levels. It therefore grows in a variety of different habitats and environments. It has wide range of medicinal and pharmacological applications. According to the Ayurveda, *C.rotundus* rhizomes are considered astringent, diaphoretic, diuretic, analgesic, antispasmodic, aromatic, carminative, antitussive, emmenagogue, litholytic, sedative, stimulant, stomachic, vermifuge, tonic and antibacterial. This paper provides review on medicinal uses and various pharmacological properties of *C.rotundus* rhizome.

Index Terms- Anti-inflammatory, Anti-pyretic, Anti-malarial, *Cyperus rotundus*, Nut grass.

I. INTRODUCTION

Herbal medicine is a major component in all traditional medical systems, and a common element in Siddha, Ayurvedic, Homeopathic, Naturopathic, Traditional Chinese medicine, and Native American medicine. Plant materials are used throughout developed and developing countries as home remedies, over-the-counter drug products and raw materials for the pharmaceutical industry, and represent a substantial proportion of the global drug market. A perfect example of medicinal plant credited with innumerable medicinal qualities validated by modern science and used since ancient times is *C.rotundus* Linn. Family - *Cyperaceae* are the largest family in the monocotyledons consisting of 109 genera and approximately 5,500 species [1].

C.rotundus L., (Family-Cyperaceae), also known as purple nutsedge or nutgrass, is a common perennial weed with slender, scaly creeping rhizomes, bulbous at the base and arising singly from the tubers which are about 1-3 cm long. The tubers are externally blackish in colour and reddish white inside, with a characteristic odour. The stems grow to about 25 cm tall and the leaves are linear, dark green and grooved on the upper surface. Inflorescences are small, with 2-4 bracts, consisting of tiny flowers with a red-brown husk. The nut is three-angled, oblong-ovate, yellow in colour and black when ripe. *C.rotundus* is indigenous to India, but are now found in tropical, subtropical and temperate regions [2].

In Asian countries, the rhizomes of *C. rotundus*, which are used as traditional folk medicines for the treatment of stomach and bowel disorders, and inflammatory diseases, have been widely investigated [3-5]. *C. rotundus* is a traditional herbal medicine used widely as analgesic, sedative, antispasmodic, antimalarial, stomach disorders and to relieve diarrhoea [6-7]. The tuber part of *C. rotundus* is one of the oldest known medicinal plants used for the treatment of dysmenorrhoeal and menstrual irregularities [8-9]. Infusion of this herb has been used in pain, fever, diarrhoea, dysentery, an emmenagogue and other intestinal problems [10]. It is a multipurpose plant, widely used in traditional medicine around the world to treat stomach ailments, wounds, boils and blisters [11-14].

A number of pharmacological and biological activities including anti-*Candida*, anti-inflammatory, antidiabetic, antidiarrhoeal, cytoprotective, antimutagenic, antimicrobial, antibacterial, antioxidant, cytotoxic and apoptotic, anti-pyretic and analgesic activities have been reported for this plant [15-24].

Previous phytochemical studies on *C.rotundus* revealed the presence of alkaloids, flavonoids, tannins, starch, glycosides and furochromones, and many novel sesquiterpenoids [25- 29].

Edible Parts: Rhizome

II.MEDICINAL USES

According to the Ayurveda, *C. rotundus* rhizomes are considered astringent, diaphoretic, diuretic, analgesic, antispasmodic, aromatic, carminative, antitussive, emmenagogue, litholytic, sedative, stimulant, stomachic, vermifuge, tonic and antibacterial.

It may be a good remedy for indigestion in the light of constituents present in it, for example, there are many enzymes for carbohydrates and minerals which act as catalyst for various biochemical reactions and helps indigestion. It is also useful for dietary management of psychotic diseases and metabolic disorders [30].

They are used in treatment of Nausea and vomiting, dyspepsia, colic, flatulence, diarrhoea, dysentery, intestinal parasites, fever, malaria, cough, bronchitis, renal and vesical calculi, urinary tenesmus, skin diseases, wounds, amenorrhoea, dysmenorrhoea, deficient lactation, loss of memory, insect bites, food poisoning, indigestion, nausea, dysuria, bronchitis, infertility, cervical

cancer and menstrual disorders, and the aromatic oils are made of perfumes and splash [31-35].

III. CHEMICAL CONSTITUENTS

Several chemical compounds have been isolated from world's worst weed *C. rotundus* [36] and some of these chemicals possess medicinal properties and are used in Latin America, China, India and elsewhere [37-39]. Various preparations of *C. rotundus* have been used for centuries in perfumes, spices and traditional medicines in India, China, Arab and Africa. It is also an important ingredient of anti-aging Ayurvedic nutraceutical Chyavanprash [39].

Different phytochemical studies on *C. rotundus* revealed the presence of alkaloids, flavonoids, tannins, starch, glycosides, furochromones, monoterpenes, sesquiterpenes, sitosterol, fatty oil containing a neutral waxy substance, glycerol, linolenic, myristic and stearic acids [25, 29, 40-41]. The major compounds isolated from essential oil and the extracts of *C. rotundus* rhizome are Alpha-cyperone, Alpha-rotunol, Beta-cyperone, Beta-pinene, Beta-rotunol, Beta-selinene, Calcium, Camphene, Copaene, Cyperene, Cyperenone, Cyperol, Cyperolone Cyperotundone D-copadiene, D-epoxyguaiene, D-fructose, D-glucose, Flavonoids, Gamma-cymene, Isocyperol, Isokobusone, Kobusone, Limonene, Linoleic-acid, Linolenic-acid, Magnesium, Manganese, C. rotunduskone, Myristic-acid, Oleanolic-acid, Oleanolic-acid-3-o-neohesperidoside, Oleic-acid, P-cymol, Patchoulone, Pectin, Polyphenols, Rotundene, Rotundenol, Rotundone, Selinatriene, Sitosterol, Stearic-acid, Sugeonol, Sugetriol [42-45].

C. rotundus contains an essential oil that provides for the characteristic odour and taste of the herb, comprised mostly sesquiterpene hydrocarbons, epoxides, ketones, monoterpenes and aliphatic alcohols. Sesquiterpenes include selinene, isocurcumenol, nootkatone, aristolone, isorotundene, cypera-2,4(15)-diene, and norrotundene, as well as the sesquiterpene alkaloids rotundines A-C. Other constituents include the ketone cyperadione, and the monoterpenes cineole, camphene and limonene. *C. rotundus* has also been shown to contain miscellaneous triterpenes including oleanolic acid and sitosterol, as well as flavonoids, sugars and minerals [44-45].

The chemical composition of the volatile oils of *C. rotundus* has been extensively studied and four chemotypes (H-, K-, M- O-types), of the essential oils from different parts of Asia have been reported [46-52].

The H-type from Japan was found to contain α -cyperone (36.6%), β -selinene (18.5%), cyperol (7.4%) and caryophyllene (6.2%). The M-type from China, Hong Kong, Japan, Taiwan and Vietnam had α -cyperone (30.7%), cyperotundone (19.4%), β -selinene (17.8%), cyperene (7.2%) and cyperol (5.6%). The O-type from Japan, Taiwan, Thailand, Hawaii and the Philippines was characterized by cyperene (30.8%), cyperotundone (13.1%) and β -elemene (5.2%). In addition, the Hawaiian O-type had cyperotundone (25.0%) and cyperene (20.7%) as the major compounds. Finally, the K-type, also from Hawaii, was

dominated by cyperene (28.7%), cyperotundone (8.8%), patchoulanyl acetate (8.0%) and sugeonyl acetate (6.9%) [47-48].

IV. PHARMACOLOGICAL ACTIVITIES

Anti Inflammatory Activity

The alcoholic extract (70% alcohol) possessed anti inflammatory activity against carrageenan induced oedema and also found effective against formaldehyde induced arthritis in albino rats [53]. In another study the petroleum ether extract of the rhizomes showed anti-inflammatory activity against carrageenan induced oedema in albino rats. The triterpenoid obtained by chromatographic separation from petroleum ether extract revealed a high potent anti-inflammatory activity. This terpenoid was also found to possess significant antipyretic and analgesic effects similar to acetyl salicylic acid. *C. rotundus* has also reported as protective in inflammatory bowel disease.

In addition, the extract suppressed the production of O₂- by phorbol ester stimulated RAW 264.7 cells in dose- and time-dependent manners. Collectively, these results suggest that the methanol extract of rhizomes of *C. rotundus* could be developed as anti-inflammatory candidate for the treatment of inflammatory diseases mediated by overproduction of NO and O₂ [54].

Another study on alcoholic extract of *C. rotundus* showed highly significant ($P < 0.001$) anti-inflammatory activity against the exudative and proliferative phases of inflammation in two animal models (carrageenan induced oedema and formaldehyde induced arthritis in rats). Its anti-inflammatory relative effect was higher than that of hydrocortisone (75.9% versus 47.3% in carrageenan-induced oedema model; 55.1% versus 35.6% in formaldehyde induced arthritis model [27, 55-57].

Antipyretic activity

The alcoholic extract of *C. rotundus* showed highly significant ($P < 0.001$) antipyretic activity against pyrexia produced in albino rats by the subcutaneous injection of suspension of dried Brewer's yeast in gum acacia in normal saline. A specific fraction obtained by chromatographic method from the petroleum ether extract was found to possess a significant antipyretic effect similar to acetyl salicylic acid when used on the same animal model [58].

Analgesic activity

The petroleum ether extract and essential oil of *C. rotundus* are reported to possess analgesic activity [58-59].

Tranquilizing activity

The ethanolic extract of *C. rotundus* showed potent tranquilizing activity in various tests: reduced the spontaneous motor activity, potentiated the pentobarbital narcosis and deranged the motor coordination, abolished the conditioned avoidance response in animals [55].

Anticonvulsant activity

Pretreatment with ethanolic extract of *C. rotundus* caused significant protection against strychnine and leptazol-induced convulsions in mice [60].

The ethanol extract of rhizomes (100mg/kg, p.o.) reduced hind limb extension and duration of convulsion significantly, ($p < 0.001$) which was comparable to standard drug Phenytoin (25mg/kg, i.p.) and Diazepam (4mg/kg, i.p.), respectively. These results suggest that the ethanol extract of its rhizomes is worthwhile to develop the potent phytoconstituent for treatment of epilepsy and the flavonoids present in ethanol extract could be attributed for anticonvulsant activity [61].

Anti-emetic activity

The ethanolic extract of *C. rotundus* in the dose of 128.1 ± 11.6 mg/kg was found to protect 50% dogs against apomorphine induced vomiting [55].

Antispasmodic activity

Ethanolic extract of *C. rotundus* produced relaxation of rabbit ileum and spasmolytic effect against contractions induced by acetylcholine, barium chloride and 5-hydroxytryptamine, showing a direct relaxant action on the smooth muscle [55].

Inhibition of gastric motility activity

The rhizome of *C. rotundus* Linn. was assessed for its cytoprotective effects against ethanol induced gastric damage. Decoctions of Rhizoma Cyperi were given orally to rats 30 min. before ethanol was administered. The findings in this study suggest that the protective action of *C. rotundus* Linn. is related to its inhibition of gastric motility and endogenous prostaglandins may play an important role [62].

Gastroprotective activity

C. rotundus extract protected against gastric mucosal injury induced by ischemia and reperfusion in rats. The mean ulcer index of rats treated with 200 and 100 mg/kg *C. rotundus* were significantly lower than that of control. The activities of glutathione-peroxidase and malondialdehyde were significantly affected by treatment of *C. rotundus* [64]. Cytoprotective effects of *C. rotundus* have been mentioned also in case of ethanol induced gastric damage in rats. Decoctions of Rhizoma Cyperi were given orally (1.25, 2.5, 4.0 g crude drug/kg) to rats 30 min before ethanol showed an ulcer inhibitory effect in a dose dependent manner. Pretreatment of rats with indomethacin (5 mg/kg) significantly reduced the gastric protective action of *C. rotundus*. The authors suggested that the gastroprotective action of *C. rotundus* is related to its inhibition of gastric motility and endogenous prostaglandins [62].

Antidiarrhoeal Activity

The methanol extract of *C. rotundus* rhizome, given orally at the doses of 250 and 500 mg/kg showed significant antidiarrhoeal activity in castor oil induced diarrhoea in mice. Among the fractions, tested at 250 mg/kg, the petroleum ether fraction and residual methanol fraction were found to retain the activity, the latter being more active as compared to the control. The ethyl acetate fraction did not show any antidiarrhoeal activity [2].

Haemodynamic (hypotensive) activity

The alcoholic extract of *C. rotundus* produced gradual and persistent fall in blood pressure and stimulated the respiration. The responses of epinephrine and acetylcholine on blood

pressure were not altered by the extract, but that of histamine was partially blocked [55].

Hypolipidaemic Activity

Wistar rats weighing 250-300 g were selected for the study. Animals were divided into 7 groups, each group comprising of 6 rats. Rats in the group 1 received normal pellet diet and received 0.1% sodium CMC solution and served as vehicle control. The rats belonging to remaining 6 groups received high fat diet for the entire duration of the study that is for 25 days. High fat diet induced hyperlipidaemia is one of the common methods to induce hyperlipidaemia. Hence hyperlipidaemia was induced by oral feeding of high fat diet. The high fat diet was comprised of the chow enriched with high calorie and 1% cholesterol. After 10 days induction of hyperlipidaemia group 2 of animals was left untreated and served as high fat diet control. The rest of the groups received following treatment for 15 days. Group 3 and group 4 treated orally with the standard drugs Simvastatin (5 mg/kg/day) and Fenofibrate (20 mg/kg/day) respectively. Groups 5, 6, 7 treated orally with aqueous extract at dose level of 100 mg/kg/day, 200 mg/kg/day, 400 mg/kg/day respectively. All the drugs were suspended in 0.1% Na CMC (vehicle). Blood samples were withdrawn from retro orbital plexus after overnight fasting. Serum was separated from blood by centrifugation for ten minutes at three thousand rpm, subsequently analyzed for total cholesterol, triglycerides and HDL cholesterol using commercially available kits (Erba Diagnostics Germany). The serum LDL was calculated by Friedwald's formula [65].

In another study administration of *C. rotundus* extract restored the age associated change in serum lipids (total cholesterol, LDL cholesterol, DL cholesterol, triglycerides and VLDL triglyceride level) to the level of young control rats. In young rats, treatment of *C. rotundus* significantly increased HDL cholesterol level [66].

Hepatoprotective activity

Ethyl acetate extract and two crude fractions, solvent ether and ethyl acetate, of the rhizomes of *C. rotundus* (Cyperaceae) were evaluated for hepatoprotective activity in rats by inducing liver damage by carbon tetrachloride. The ethyl acetate extract at an oral dose of 100 mg/kg exhibited a significant protective effect by lowering serum levels of glutamic oxaloacetic transaminase, glutamic pyruvic transaminase, alkaline phosphatase and total bilirubin. These biochemical observations were supplemented by histopathological examination of liver sections. Silymarin was used as positive control [67].

Inhibitory activity on Brain Na⁺/K⁺-ATP-ase

Extract of *C. rotundus* showed high potent inhibitory activity on crude enzyme Na⁺/K⁺-ATP-ase from rat brain [68].

Anti-obesity activity

C. rotundus preparations (powder in fine suspension, aqueous and alcoholic extracts) exhibited a lipolytic action and mobilized fat from the adipose tissues in rats, thus helping to reduce the obesity [69].

A pilot study carried out on 30 obese people who were administered the powdered tuber of *C. rotundus* for 90 days,

showed reduction in weight along with a decrease in serum cholesterol and triglycerides [70].

Antiarthritic activity

Singh and his co-workers were first to discover anti-inflammatory, anti-pyretic and anti-rheumatic activity of *C. rotundus* [27,29,35]. A double blind trial of crude powder of *C. rotundus*, *Withania somnifera* and their combination (1:1) was carried out in 200 patients suffering from rheumatoid arthritis. Out of the 200 patients selected for the study 196 completed the trial of 3 months. Each group (including placebo group) consisted of 50 patients. Each patient received 500 mg capsule three times a day for three months. During this period biweekly general assessment based on global criteria (duration of morning stiffness, grip strength, articular index, consumption of escape analgesic, erythrocyte sedimentation rate, haemoglobin, rheumatoid factor titre, x-ray findings) was made. *C. rotundus* was more effective than *W. somnifera*, and when both drugs were combined, the response was better than the response of single drug. Also the patients' preference (against escape analgesic) was highest in the case of combined herbs [55-56,58, 71-72].

Wound healing activity

An alcoholic extract of tuber parts of *C. rotundus* was examined for wound healing activity in the form of ointment in three types of wound models on rats: the excision, the incision and dead space wound model. The extract ointments showed considerable difference in response in all the above said wound models as comparable to those of a standard drug nitrofurazone ointment (0.2 % w/w NFZ) in terms of wound contracting ability, wound closure time and tensile strength [73].

Antioxidant activity

A combination of spices (*Piper nigrum*, *Piper longum* and *Zingiber officinale*), herbs (*Cyperus rotundus* Linn. and *Plumbago zeylanica*) and salts make up Amrita Bindu. The study was focused to evaluate the antioxidant property of individual ingredients in Amrita Bindu against the free radical 2,2'-azinobis-(3-ethylbenzothiazoline-6-sulphonic acid) (ABTS). The analysis revealed the antioxidant potential of the ingredients in the following order: *Piper nigrum* > *Piper longum* > *Cyperus rotundus* > *Plumbago zeylanica* > *Zingiber officinale*. These results reveal that Amrita Bindu, a salt-spice-herbal mixture containing *C. rotundus* Linn. exerts a promising antioxidant potential against free radical induced oxidative damage [74].

Anticancer activity

Anticancer *C. rotundus* ethanolic extract was found to have only weak to moderate anticancer activity (LC50=2.528-4.939 mg/ml calculated from dose-dependent cell death) in a study which used neuro-2a cells for screening of plants with tumoricidal effects [75]. Another study showed that *C. rotundus* essential oil was very effective against L1210 leukaemia cells line. This result correlated with significantly increased apoptotic DNA fragmentation [21].

Antidiabetic activity

Oral daily administration of 500 mg/kg of the extract (once a day for seven consecutive days) significantly lowered the blood glucose levels in rats with alloxan induced diabetes. The scientists concluded that this antihyperglycemic activity can be attributed to its antioxidant activity as *C. rotundus* showed a strong 1,1-diphenyl-2-picrylhydrazyl (DPPH) radical scavenging action *in vitro*. These results are convergent with *C. rotundus* potential to suppress AGE formation and protein oxidation in a model of fructose-mediated protein glycoxidation. Scientists concluded that, since non-enzymatic glycation has been shown to correlate with severity of diabetes and its complications, *C. rotundus* could be a candidate for targeting diabetic complications [17, 76].

Antimicrobial activity

In-vitro antimicrobial activity by agar disc diffusion and agar well diffusion method was evaluated for aqueous and ethanolic extracts. The ethanolic extract was active against all the investigated bacterial strains, while aqueous extract was inactive. In another study acetone and ethanol extracts showed significant broad spectrum antibacterial activity in disc diffusion method [9, 77]. Antimicrobial activity tests were carried out on human pathogens bacteria (gram negative and gm positive) and fungi viz. *C. albicans* and *A. niger*. The highest percentage of inhibition was observed against *K. pneumoniae* (133.33%). Amoxicillin 20µg/ml and ethanol (as fungicide) 70% were used as positive control. Moderate inhibition was observed in case of *A. niger* and *S. aureus* (90 and 70% respectively). No zone of inhibition was observed in *Acinetobacter* and *Candida*.

Antibacterial Activity

The oil of *C. rotundus* showed a remarkable activity against gram-positive bacteria *Staphylococcus aureus* and *Enterococcus faecalis* [73,78]. Another study stated that a marked inhibitory effect of *C. rotundus* was observed against *Salmonella enteritidis*, *Staphylococcus aureus* and *Enterococcus faecalis* with total oligomers flavonoids (TOFs) and ethyl acetate extracts [79, 80].

Antimalarial Activity

Activity guided investigation of sesquiterpenes *C. rotundus* rhizomes showed *in-vitro* antimalarial activity against *Plasmodium falciparum* [81]. Some Tanzanian medicinal plants were extracted and tested for *in vitro* antimalarial activity, using the multidrug resistant K1 strain of *Plasmodium falciparum*. Of the forty-nine plants investigated, extracts of three plants were found to have an IC50 between 5-10 mg/ml; extracts of 18 other plants showed an IC50 between 10 and 50 mg/ml, all others were less active. The three most active extracts were obtained from the tubers of *C. rotundus* Linn. the root bark of *Hoslundia opposita* Vahl. and the root bark of *Lantana camara* L [6].

The underground parts of several weedy species contain essential oils, about 0.5-1% in the case of the fresh tubers of *C. rotundus*, mainly consisting of terpenoids or sesquiterpenoids (e.g. cyperone, cyperol, cyperolone, cyperene, copadiene, epoxyguaiaene, rotundone, rotundol, patchoulone (cyperotundon), kobusone, sugeonolacetate, sugetriol, oxido-eudesmenol, C.

rotunduskone and 'BETA'-selinene).When Tanzanian medicinal plants were screened, *C. rotundus* showed activity in a test for in vitro antimalarial activity [82].

Ovicidal and larvicidal activities

The ovicidal and larvicidal efficacy of essential oils extracted from the tubers of *Cyperus giganteus* and *Cyperus rotundus* Linn. was studied on eggs and fourth instar larvae of *Aedes albopictus*. The eggs and larvae were exposed to serial concentration of the oils ranging from 5-150 ppm and kept under observation for 24h. Both the oils showed remarkable ovicidal and larvicidal activities indicated by EC50 values of <5 ppm and LC50 and LC90 values of <20 ppm. The results obtained suggest that the essential oils of these *Cyperus* species can serve as a potential source of natural mosquitocidal agents [83]

Anti Candida activity,

Essential oils and alcoholic extracts from the leaves and/or roots of 35 medicinal plants commonly used in Brazil were screened for anti *Candida albicans* activity. Essential oils from 13 plants showed anti *Candida* activity, including *Aloysia triphylla*, *Anthemis nobilis*, *Cymbopogon martini*, *Cymbopogon winterianus*, *Cyperus articulatus*, ***Cyperus rotundus* Linn.**, *Lippia alba*, *Mentha arvensis*, *Mikania glomerata*, *Mentha piperita*, *Mentha sp.*, *Stachys byzantina*, and *Solidago chilensis*. The ethanol extract was not effective at any of the concentrations tested. Chemical analyses showed the presence of compounds with known antimicrobial activity, including 1,8-cineole, geraniol, germacrene-D, limonene, linalool, and menthol [84].

Cytoprotective effects

The rhizome of *C. rotundus* was assessed for its cytoprotective effects against ethanol induced gastric damage. Decoctions of Rhizoma Cyperi were given orally (1.25, 2.5, 4.0 g crude drug/kg) to rats 30 min before ethanol (40% v/v, 10mL/kg) was administered. The decoction showed an ulcer inhibitory effect in a dose dependent manner. Moreover, the activity was also observed when the decoction was given subcutaneously (0.3-0.6 g/kg), suggesting that the herb possessed systemic effects on protecting the stomach. Compared with controls, gastric motility of the ethanol-treated rats was delayed significantly by either oral (2.5-4.0 g/kg) or subcutaneous (0.3g/kg) administration of the decoction. Pretreatment of rats with indomethacin (5 mg/kg) significantly reduced the gastric protective action of *C. rotundus* [85].

Toxicological studies

Rats were divided into two groups of ten animals (five males, five females). The ethanol extract (2,500 mg/ml in 10% dimethylsulfoxide, DMSO) was orally administered to rats at a single dose of 5,000 mg/kg body weight, while the control group received only vehicle. The animals were monitored for the appearance of toxicity signs over 14 days. The animals that died within this period were necropsied. All rats were weighed and sacrificed on the 14th day following administration. Finally, the vital organs including heart, lungs, livers, kidneys, spleen, adrenals, sex organs and brain were grossly examined.

In the acute toxicity test at the dose of 5,000 mg/kg, all rats did not exhibit signs of toxicity and mortality after a single oral administration of 95% ethanol extract from the rhizomes of *C. rotundus*. Results of the subacute toxicity showed that administration of the ethanol extract from the rhizomes of *C. rotundus* at a dose of 1,000 mg/kg daily over 14 days did not cause mortality or behavioral changes [86].

Another study for the purpose of the test, in bred wistar strain rats (250-300 g) of both sexes were selected. The animals were housed in polypropylene cages (6rats per cage) under good hygienic conditions natural light / dark cycle. The animals were given free access to standard pellet diet and water. The acute toxicity study was carried out as per OECD guideline (OECD/OCDE 423 OECD Guideline for testing of chemicals Acute Oral Toxicity –Acute Toxic Class Method Adopted: 17th December 2001). Thus the oral acute toxicity tests revealed that the extract of *C.rotundus* rhizomes was safe up to the administered dose 2000 mg/kg.

Another acute toxicological studies showed no mortality or morbidity up to 2000mg/kg body weight in Wistar rats. Sub chronic toxicity study revealed that, food, water consumption and body weight of animals didn't vary significantly. But the hematological parameters showed an increase in WBC count and Hemoglobin level. The kidney function and liver function didn't change even after long term exposure [87].

V. DISCUSSION

C. rotundus Linn., commonly known as nut grass and locally. It is said to possess antidiarrheal, anti-inflammatory and antipyretic activities. The tubers are used in Ayurvedic medicine and have been mentioned in ancient texts for various ailments. Some studies have reported antidiarrheal activity of *C.rotundus*. Antidiarrheal action in castor oil-induced diarrhoea and in irritable bowel syndrome in animal models has been demonstrated. Previous studies with the essential oil of *C. rotundus* showed it to be more bactericidal against Gram-positive bacteria.

The major constituents present in *C.rotundus* are essential oil, triterpenes, polyphenol, alkaloids and flavonoids. However, none of these have been attributed with antidiarrheal activity.

The decoction used showed the presence of carbohydrates, reducing sugars, proteins, amino acids, tannins, flavonoids and saponins. Tannins and flavonoids, in general, have been reported to have antidiarrheal activity.

This study shows that *C.rotundus* has limited antimicrobial action and have. *C. rotundus* with a large number of biologically active phytochemicals has diverse variety of pharmacological properties, as described above, has been found effective in the treatment of chronic disorders. Its therapeutic effects are excellent and no adverse reaction was observed.

VI. CONCLUSION

The above collected information suggest that *C. rotundus* has limited activity against different forms of infectious diarrhoea due to its selective activity against diarrheal pathogens. Traditional uses of natural compounds, especially of plant origin received much attention as they are well tested for their efficacy and generally believed to be safe for human use. Thorough screening of literature available on *C. rotundus* depicted the fact that it is a popular remedy among the various ethnic groups, Ayurvedic and traditional practitioners for treatment of ailments. Researchers are exploring the therapeutic potential of this plant as it has more therapeutic properties which are not known.

REFERENCES

- Govaerts R, Simpson DA, Goetghebeur P, Wilson K, Egorova T, Bruhl JJ (2007) World checklist of Cyperaceae. The Board of Trustees of the Royal Botanic Gardens, Kew. Available at <http://www.kew.org/wcsp/monocots/>, Accessed on 1 October 2007.
- Uddin SJ, Mondal K, Shilpi JA, Rahnan MT. Antidiarrhoeal activity of *Cyperus rotundus*. *Fitoterapia* 2006; 77 (2): 134–13
- Dang GK, Parekar RR, Kamat SK, Scindia AM, Rege NN., Antiinflammatory activity of *Phyllanthus emblica*, *Plumbago zeylanica* and *Cyperus rotundus* in acute models of inflammation., *Phytother Res.* 2011Jun;25(6):904-8. doi: 10.1002/ptr.3345. Epub 2010 Dec 3.
- Gupta MB, Palit TK, Singh N, Bhargava KP. Pharmacological studies to isolate the active constituents from *Cyperus rotundus* possessing anti-inflammatory, anti-pyretic and analgesic activities. *Indian Journal of Medical Research* 1971; 59: 76–82.
- Won-Gil Seo, Hyun-Ock Pae, Gi-Su Oh, Kyu-Yun Chai, Tae-Oh Kwon, Young-Gab Yun, Na-Young Kim, Hun-Taeg Chung, Inhibitory effects of methanol extract of *Cyperus rotundus* rhizomes on nitric oxide and superoxide productions by murine macrophage cell line, RAW 264.7 cells, *Journal of Ethnopharmacology*, Volume 76, Issue 1, June 2001, Pages 59–64.,
- Weenen H, Nkunya MH, Bray DH, Mwasumbi LB, Kinabo LS, Kilimali VA. Antimalarial activity of Tanzanian medicinal plants. *Planta Medica* 1990a; 56: 368–370.
- Zhu M, Luk HH, Fung HS, Luk CT. Cytoprotective effects of *Cyperus rotundus* against ethanol induced gastric ulceration in rats. *Phytother. Res* 1997; 11: 392–394.
- Yu J., Lei G., Cai L. and Zou Y. “Chemical composition of *C. rotundus* extract”. (2004). *J. Phytochemistry*. 65: 881-89.
- Zeid Abdul-Majid Nima, Majid Sakhi Jabier, Raghidah Ismaeel Wagdi, Huda Abd Al-Kareem Hussain., Extraction, Identification and Antibacterial activity of *Cyperus* oil from Iraqi *C rotundus*., *Eng.& Technology*, Vol.26, No.10, 2008.
- Umerie SC, Ezeuzo HO. Physicochemical characterization and utilization of *Cyperus rotundus* starch. *Bioresour. Technol* 2000; 72: 193–196
- Oliver-Bever, B. *Medicinal Plants in Tropical West Africa*; Cambridge University Press: Cambridge, UK, 1986; p. 200.
- Puratuchikody, A.; Nithya, D.C.; Nagalakshmi, G. Wound Healing Activity of *Cyperus rotundus* Linn. *Indian J. Pharm. Sci.* 2006, 68, 97-101.
- Joshi, A.R.; Joshi, K. Indigenous knowledge and uses of medicinal plants by local communities of the Kali Gandaki Watershed Area, Nepal. *J. Ethnopharmacol.* 2000, 73, 175-183.
- El-Kamali, H.H.; El-Khalifa, K.F. Folk medicinal plants of riverside forests of the Southern Blue Nile district, Sudan. *Fitoterapia* 1999, 70, 493-497.
- Durate, M.C.T.; Figueira, G.M.; Sartoratto, A.; Rehder, V.L.G.; Delarmelina, C. Anti-Candida activity of Brazilian medicinal plant. *J. Ethnopharmacol.* 2005, 97, 305-311.
- Sundaram, M.S.; Sivakumar, T.; Balamurugan, G. Anti-inflammatory effect of *Cyperus rotundus* Linn. Leaves on acute and subacute inflammation in experimental rat models. *Biomedicine* 2008, 28, 302-304.
- Raut, N.A.; Gaikwad, N.J. Antidiabetic activity of hydro-ethanolic extract of *Cyperus rotundus* in alloxan induced diabetes in rats. *Fitoterapia* 2006, 77, 585–588.
- Kilani, S.; Ben Ammar, R.; Bouhleb, I.; Abdelwahed, A.; Hayder, N.; Mahmoud, A.; Ghedira, K.; Chekir-Ghedira, L. Investigation of extracts from (Tunisian) *Cyperus rotundus* as antimutagens and radical scavengers. *Environ. Toxicol. Pharmacol.* 2005, 20, 478-484.
- Zhu, M.; Luk, H.H.; Fung, H.S.; Luk, C.T. Cytoprotective effects of *Cyperus rotundus* against ethanol induced gastric ulceration in rats. *Phytother. Res.* 1997, 11, 392 -394.
- Kilani, S.; Bouhleb, I.; Ben Ammar, R.; Ben Sghair, M.; Skandrani, I.; Boubaker, J.; Mahmoud, A.; Dijoux-Franca, M.G.; Ghedira, K.; Chekir-Ghedira, L. Chemical investigation of different extracts and essential oil from the tubers of (Tunisian) *Cyperus rotundus*. Correlation with their antiradical and antimutagenic properties. *Ann. Microbiol.* 2007, 57, 657-664.
- Kilani, S.; Ledauphin, J.; Bouhleb, I.; Ben Sghaier, M.; Boubaker, J.; Skandrani, I.; Mosrati, R.; Ghedira, K.; Barillier, D.; Chekir-Ghedira L. Comparative study of *Cyperus rotundus* essential oil by a modified GC/MS analysis method. Evaluation of its antioxidant, cytotoxic, and apoptotic effects. *Chem. Biodivers.* 2008, 5, 729-742.
- Dhillon, R.S.; Singh, S.; Kundra, S.; Basra, A.S. Studies on the chemical composition and biological activity of essential oil from *Cyperus rotundus* Linn. *Plant Growth Regul.* 1993, 13, 89-93.
- Pal, D.K.; Dutta, S. Evaluation of the Antioxidant activity of the roots and Rhizomes of *Cyperus rotundus* L. *Indian J. Pharm. Sci.* 2006, 68, 256-258.
- Neffatti, A.; Ben Ammar, R.; Dijoux-Franca, M.G.; Ghedira, K.; Chekir-Ghedira, L. *In vitro* evaluation of antibacterial, antioxidant, cytotoxic and apoptotic activities of the tubers infusion and extracts of *Cyperus rotundus*. *Bioresour. Technol.* 2008, 99, 9004 9008.
- Harborne, J.B.; Williams, C.A.; Wilson, K.L. Flavonoids in leaves and inflorescences of Australian *Cyperus* species. *Phytochemistry* 1982, 21, 2491-2507.
- Umerie, S.C.; Ezeuzo, H.O. Physicochemical characterization and utilization of *Cyperus rotundus* starch. *Bioresour. Technol.* 2000, 72, 193-196.
- Kapadia, V.H.; Naik, V.G.; Wadia, M.S.; Dev, S. Sesquiterpenoids from Essential oil of *Cyperus rotundus*. *Tetrahedron Lett.* 1967, 4661.
- Trivedi, B.; Motl, O.; Herout, V.; Sorm, F. Composition of the oil from *Cyperus rotundus*: Structure of patchoulone. *Coil. Czech. Chem. Commun.* 1984, 29, 1675-1688.
- Sri Ranjani, S.; Prince, J.; Physico-chemical and Phyto-chemical study of rhizome of *Cyperus rotundus* Linn. *International Journal of Pharmacology and Pharmaceutical Technology (IJPT)*, ISSN: 2277 – 3436, Volume-1, Issue- 2, 2012. 42-46.
- Anonymous. 1950. *The Wealth of India : Raw Materials*. II. Publications and Information Directorate, C.S.I.R., New Delhi.
- Yeung, Him-Che. *Handbook of Chinese Herbs and Formulas*. Institute of Chinese Medicine, Los Angeles 1985
- Duke, J. A. and Ayensu. E. *S.Medicinal Plants of China* Reference Publications, Inc. 1985 ISBN 0-917256-20-4
- Bown, D. *Encyclopaedia of Herbs and their Uses*. Dorling Kindersley, London. 1995 ISBN 0-7513-020-31
- Chopra. R. N., Nayar, S. L. and Chopra. I. C. *Glossary of Indian Medicinal Plants (Including the Supplement)*. Council of Scientific and Industrial Research, New Delhi. 1986
- Medicinal Plants in the Republic of Korea* World Health Organisation, Manila 1998 ISBN 92 9061 120 0
- Sonwa, M.M.; Koenig, W.A. Chemical study of essential oil *Cyperus rotundus*. *Phytochemistry* 2001, 58, 799-810.
- Ellison, C.A. & R.W. Barreto. 2004. Prospects for the management of invasive alien weeds using co-evolved fungal pathogens: a Latin American perspective *Biological Invasions* 6: 23-45.
- Gupta, M. B., Palit, T. K., Signh, N., and Bhargava, K. P., Pharmacological studies to isolate the active constituents from *Cyperus rotundus* possessing anti-inflammatory, anti-pyretic and analgesic activities. *Indian J. Med. Res*, 59, 76-82 (1971)
- Sharma, R. & Gupta, R. 2007. *Cyperus rotundus* extract inhibits acetylcholinesterase activity from animal and plants as well as inhibits germination and seedling growth in wheat and tomato, *Life Sciences* 80: 2389-2392.
- Akperbekova B A, Pharmacognostic study of the cyperus retundus rhizome. *Farmatsiya*, 1967, 16(1), 43-45
- Dutta S C and Mukerji B, Pharmacognosy of Indian Root and rhizome drugg, Manager of Publications Delhi, 1949, Vol.148, 135-136.
- Salman Khan, Ran Joo Choi, Dong Ung Lee, Yeong Sik Kim, Sesquiterpene derivatives isolated from *Cyperus rotundus* L., inflammatory signaling mediated by NF κ B, *Natural Product Sciences*, 2011, 17(3), 250-255.

43. Oladipupo A. Lawal and Adebola O. Oyedeji, Chemical Composition of the Essential Oils of *Cyperus rotundus* L. from South Africa, *Molecules* 2009; 14: 2909-2917
44. Sonwa MM, König WA Chemical study of the essential oil of *Cyperus rotundus* Phytochemistry. 2001 Nov;58(5):799-810.
45. Jeong, S.J.; Miyamoto, T.; Inagaki, M.; Kim, Y.C.; Higuchi, R. Rotundines A-C, three novel sesquiterpene alkaloids from *Cyperus rotundus*. *J. Nat. Prod.* **2000**, *63*, 673-675.
46. Kilani, S.; Ledauphin, J.; Bouhlel, I.; Ben Sghaier, M.; Boubaker, J.; Skandrani, I.; Mosrati, R.; Ghedira, K.; Barillier, D.; Chekir-Ghedira L. Comparative study of *Cyperus rotundus* essential oil by a modified GC/MS analysis method. Evaluation of its antioxidant, cytotoxic, and apoptotic effects. *Chem. Biodivers.* 2008, *5*, 729-742.
47. Komai, K.; Tang, C. A Chemotype of *Cyperus rotundus* in Hawaii. *Phytochemistry* 1989, *28*, 1883-1886.
48. Komai, K.; Shimizu, M.; Tang, C.T.; Tsutsui, H. Sesquiterpenoids of *Cyperus bulbosus*, *Cyperus tuberosus* and *Cyperus rotundus*. *Mem. Fac. Agr. Kinki Univ.* 1994, *27*, 39-45.
49. Ekundayo, O.; Oderinde, R.; Ogundeyin, M.; Biskup, E.S. Essential oil constituents of *tuberosus* Rottb. Rhizomes. *Flav. Fragr. J.* 1991, *6*, 261-264.
50. Kilani, S.; Abdelwahed, A.; Chraief, I.; Ben Ammar, R.; Hayder, N.; Hammami, M.; Ghedira, K.; Chekir-Ghedira, L. Chemical composition, antibacterial and antimutagenic activities of essential oil from (Tunisian) *Cyperus rotundus*. *J. Essent. Oil Res.* 2005, *17*, 695-700.
51. Zoghbi, M.D.G.B.; Andrade, E.H.A.; Carreira, L.M.M.; Rocha, E.A.S. Comparison of the mcomponents of the essential oils of "priprioca": *Cyperus articulatus* var. *articulatusarticulatus* var. *nodosus* L., *C. prolixus* Kunth and *C. rotundus* L. *J. Essent. Oil Res* 42-46.
52. Jirovetz, L.; Wobus, A.; Buchbauer, G.; Shafi, M.P.; Thampi, P.T. Comparative analysis of the essential oil and SPME-headspace aroma compounds of *Cyperus rotundus* L. roots/tubers from South-India using GC, GC-MS and olfactometry. *J. Essent. Oil-Bearing Plants* 2004,
53. Sundaram, M.S.; Sivakumar, T.; Balamurugan, G. Anti-inflammatory effect of *Cyperus rotundus* Linn. Leaves on acute and subacute inflammation in experimental rat models. *Biomedicine* 2008, *28*, 302-304.
54. Seo WG, Pae HO, Oh GS, Chai KY, Kwon TO, Yun YG, et al. Inhibitory effects of methanol extract of *Cyperus rotundus* Linn. Linn. rhizomes on nitric oxide and superoxide productions by murine acrophage cell line, RAW 264.7 cells. *J Ethnopharmacol.* 2001; 76(1): 59-64.
55. Singh N, Kulshrestha VK, Gupta MB and Bhargava K P.. A pharmacological study of *Cyperus rotundus*, *Indian J Med, Res*, 1970,58, 103-109.
56. Singh N, Kulshrestha V K, Gupta M B and Bhargava K P, Pharmacological studies on *Cyperus rotundus* , *Indian J Pharm*,1969,1(2), 9.
57. Singh N and Gilca M, Herbal Medicine – Science embraces tradition – A new insight into the ancient Ayurveda, Lambert Academic Publishing, Germany, 2010, pp. 139-148.
58. Gupta MB, Palit TK, Singh N, Bhargava KP. Pharmacological studies to isolate the active constituents from *Cyperus rotundus* possessing anti-inflammatory, anti-pyretic and analgesic activities. *Indian Journal of Medical Research* 1971; 59: 76–82.
59. Birdar S, Kangralkar V A, Mandavkar Y, Thakur M and Chougule N, Anti-inflammatory, anti-arthritis, analgesic anticonvulsant activity of cyperus essential oils, *Int J Pharm Pharmaceut Sci*, 2010, 2(4), 112-115.
60. Pal D, Dutta S and Sarkar, A evaluation of CNS activities of ethanol extract of roots and rhizomes of *Cyperus rotundus* in mice, *Acta Poloniae Pharmaceut Drug Res.* 2009, 66(5), 535-541.
61. Shivakumar S I, Suresh H M, Hallikeri C S, Hatapakki B C, Handiganur J S, Kuber S and Shivakumar B. Anticonvulsant effect of *Cyperus rotundus* Linn. rhizomes in rats, *J Nat Rened*, 2009, 9(2), 192-196.
62. Zhu M, Luk HH, Fung HS, Luk CT. Cytoprotective effects of *Cyperus rotundus* Linn. against ethanol induced gastric ulceration in rats. *Phytother Res* 1997; 11(5): 392-94.
63. Guldur M E, Ozgonul A, Kilic I H, Sogut O and Ozaslan M, Gastroprotective effect of *Cyperus rotundus* extract against gastric mucosal injury induced by ischemia and reperfusion in rats, *Int J Pharmacol* , 2010, 6, 104-110.
64. Santhosh Kumari, Govindasamy, Sukumarb. Lipid lowering activity of *Eclipta prostrata* in experimental hyperlipidemia. *Journal of Ethnopharmacology* 2006; 105: 332–335.
65. Friedwald W. T., Levy R. I, Fredrickson D. S., Estimation of concentration of Low- Density Lipoprotein cholesterol in plasma, without use of preparative ultracentrifuge. *Clinical chemistry* 1972; 18: 499-502.
66. Nagulendran K R, Mahesh R and Begum V H, Preventive role of *Cyperus rotundus* rhizomes extract on age associated changes in glucose and lipids, *Pharmacologyonline*,2007,2, 318-325
67. Kumar S. V. S., Mishra H., Hepatoprotective Activity Of Rhizomes Of *Cyperus Rotundus* Linn Against Carbon Tetrachloride-Induced Hepatotoxicity: 2005, 67:1: 84-88
68. Ngamrojanavanish N, Manaki S and Pornpakakul S, Inhibitory activity of selected Thai medicinal plants on Na+/K+-ATP-ase, *Fitoterapia*, 2006, 77 (6), 481-483
69. Bambhole V D, Effect of some medicinal plants preparations on adipose tissue metabolism, *Ancient Sci Life* 1988, 8, 117-124
70. Karnick C R, Clinical evaluation of *Cyperus rotundus* Linn. (motha on obesity): A randomized double blind placebo controlled trial on Indian patients, *Indian Med*, 1992, 4(2),7-10.
71. Singh N and Mittal H C, In: *Medicinal Plants*, Vol. I, by V Ramalingam (Ed), MSS Information Corporation, New York, USA, 1974.
72. Singh N, Singh S P, Dixit K S, Saxena R C and Kohli R P, A placebo controlled clinical trial of *Cyperus rotundus*, *Withania somnifera* and their combination in cases of rheumatoid arthritis, *Proc International Seminar on Clinical Pharmacology in Developing Countries*, Lucknow, India, 1986, Vol. 2, pp. 18-21
73. Puratchikody A, Devi Nithya C, Nagalakshmi G. Wound healing activity of cyperus rotundus linn. *Indian journal of pharmaceutical sciences* 2006; 68: 97-101.
74. Natarajan B, Paulsen BS. An ethnopharmacological study from Thane district, Maharashtra, India: Traditional knowledge compared with modern biological science. *Pharmaceutical Biology.* 2000; 38: 139–151.
75. Mazzio E A and Soliman K F A, In vitro screening for the tumoricidal properties of international medicinal herbs, *Phytother Res*,2009, 23(3), 385-398.
76. Ardestani A and Yazdanparast R, *Cyperus rotundus* suppresses AGE formation and protein oxidation in a model of fructose-mediated protein glycooxidation, *Int J Biol Macromol* , 2007, 41(5), 572-578.
77. Singh S; SK Sharma. *Indian J. Nat. Prod.*, 2005, 21, 1, 16-17.
78. Jigna Parekh, and Sumitra Chanda., In-vitro Antimicrobial Activities of Extractsof *Launaea procumbens* Roxb. (Labiatae), *Vitis vinifera* L. (Vitaceae) and *Cyperus rotundus* L. (Cyperaceae) *African Journal of Biomedical Research*, Vol. 9, Vol. 2, May, 2006, pp. 89-93.
79. Chandratre R. S., Chandarana S, Mengi S. A., Effect of Aqueous Extract of *Cyperus rotundus* on Hyperlipidaemia in Rat Model., *International Journal of Pharmaceutical & Biological Archives* 2012; 3(3):598-600.
80. Kilani S, Ben Sghaier M, Limem I, Bouhlel I, Boubaker J, Bhourri W, Skandrani I, Neffatti A, Ben Ammar R, Dijoux-Franca M G, Ghedira K and Chekir-Ghedira L, In vitro evaluation of antibacterial, antioxidant, cytotoxic and apoptotic activities of the tubers infusion and extracts of *Cyperus rotundus*, *Bioresour Technol* , 2008, 99(18), 9004-9008.
81. Thebtaranonth, C., Thebtaranonth, Y., Wanaupathamkul,S., and Yuthavong, Y., Antimalarial sesquiterpenes from tubers of *Cyperus rotundus* : structure of 10,12-peroxyca-lamenene, a sesquiterpene endoperoxide. *Phytochemistry*, 40, 125-128 (1995).
82. Nguyen Khac Khoi, 1999. *Cyperus* L.In: de Padua, L.S., Bunyapraphatsara, N. and Lemmens, R.H.M.J. (Editors). *Plant Resources of South-East Asia* No. 12(1): Medicinal and poisonous plants 1. Backhuys Publisher, Leiden, The Netherlands, pp. 222-229.
83. KemprajVivek, Bhat Sumangala K. Ovicidal and larvicidal activities of *Cyperus giganteus* Vahl and *Cyperus rotundus* Linn. essential oils against *Aedes albopictus* (Skuse), *Natural Product Radiance* 2008; 7(5): 416-419
84. Duarte MC, Figueira GM, Sartoratto A, Rehder VL, Delarmelina C. Anti-Candida activity of Brazilian medicinal plants. *J Ethno pharmacol.* 2005; 28; 97(2): 305-11.
85. Zhu M.; Luk H. H.; Fung H. S. ; Luk C. T. Cytoprotective effects of *Cyperus rotundus* against ethanol induced gastric ulceration in rats PTR. *Phytotherapy research* ISSN 0951-418X 1997, vol. 11, n°5, pp. 392-394.
86. Thanabhorn S, Jaijoy K, Thamaree S, Ingkaninan K. Panthon A., Acute and Subacute Toxicities of the Ethanol Extract from the Rhizomes of *Cyperus rotundus* Linn., *Mahidol University Journal of Pharmaceutical Sciences* 2005; 32(1-2): 15-22.
87. Jebasingh D, Venkataraman S, Jackson D D, Emerald B S., Physiochemical and toxicological studies of the medicinal plant *Cyperus rotundus* L (Cyperaceae)., *International Journal of applied Research in natural products.*, 2012., Vol 5, No 4.

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