

# Role of Cd and Hg on biochemical contents of fennel and its reduction by exogenous treatment of nitrogen

Mohsin Anayat, Rajneesh Kumar Agnihotri, Rashmi Vamil, Sanjay Kumar and Rajendra Sharma

Department of Botany, School of Life Sciences, Khandari, Agra, Dr. B.R. Ambedkar University Agra-282002

**Abstract-** Heavy metal contamination is a serious environmental problem that limits crop production and endangers human health. The present study was conducted at Department of Botany, School of Life Sciences, Dr. B.R. Ambedkar University, Agra for assessing the toxic effect of cadmium and Mercury at different concentrations viz. 10, 20 and 50  $\mu\text{M}$  on the chlorophylls, proline and protein content of *Foeniculum vulgare* L. The standard solution was prepared using standard metal of inorganic ventures. These heavy metals affected the chlorophylls, carotenoids, protein and proline content of fennel seedlings as compared to control. Carotenoids were less affected as compared to chlorophyll 'a' and 'b' at low concentration (10 $\mu\text{M}$ ), while at higher concentrations i.e. 20 and 50 $\mu\text{M}$ , the chlorophylls, carotenoids and protein content of the seedlings were reduced drastically. However the addition of nitrogen (5 mM) minimized the effect of these heavy metals to some extent. The proline content of plants was increased under Cd and Hg treatments at all concentrations. However in this case additional supply of nitrogen in the form of Ammonium nitrate decreased proline content of plants at all concentrations of these heavy metals (Cd and Hg).

**Index Terms-** Chlorophyll, Fennel, Heavy metals, Proline and Protein

## I. INTRODUCTION

*Foeniculum vulgare* (L.) is a biennial or short lived perennial herb attaining a height of up to 2 meter. Fennel prefers loamy soil, rich in organic matter with a pH between 6.5 and 8.0 and soil temperature between 50 – 75 °F. It is a native of Mediterranean region and Europe but is commonly cultivated throughout India especially in Assam, Maharashtra, Punjab and Gujarat (Kaur and Arora, 2006). It is used as a spice and also as an important ingredient in various folklore medicines throughout the world. India is well known historically as a land of spices and aromatic plants and continues to be one of the leading producers of spices and medicinal plants in the world (Prajapati *et al.*, 2005). Spices are dried parts of the plants, which have been used as diet components often to improve colour, aroma and acceptability of food. With the current emphasis of eating healthy diets that are low in fat and salts, people are turning to various herbs and spices to flavour their food. But the overall growth and productivity of these plants have been reduced to considerable extent by the heavy metal pollution in the air, soil and water (Husain *et al.*, 1995). Toxic heavy metal interferes with several metabolic processes, causing toxicity to the plants revealed by reduced root growth and phytomass, chlorosis,

photosynthetic impairing, stunting and finally plant death (Sinha *et al.*, 2007). Among list of various heavy metals, the Cd and Hg top the relative toxicity of metals to flora and fauna. Mercury has been found to reduce phytomass, total chlorophyll, photosynthesis, nitrogen and phosphorous contents in aquatic plants. Mercury (Hg) has become a problem of current interest as a result of environmental pollution on global scale (Aliu *et al.*, 2013). Cd is a non essential heavy metal that does not have any metabolic function in plants (Bavi *et al.*, 2011). It can be incorporated and accumulated by all organisms in large amounts and disturbs physiological metabolism in plants like transpiration, photosynthesis, respiration and nitrogen assimilation (Wang *et al.*, 2008). Proteins are the main components of nucleic acid, cell membrane and other cell organelles. Heavy metals are known to reduce protein content of various plants (Bavi *et al.*, 2011; Balestrasse *et al.*, 2003). Most common form of nitrogen i.e.

$\text{NO}_3^-$  are highly reactive and mobile and are generally susceptible to losses to heavy metal stress condition.  $\text{NH}_4^+$ , which are water soluble and are easily available for absorption by plants.

## II. MATERIAL AND METHODS

The seeds of *Foeniculum vulgare* L. were obtained from National Seed Corporation, Sikandara, Agra. Seeds with uniform size, colour and weight were chosen for the experimental purpose. Two types of experiments – petridish experiment and pot and sand culture experiment were conducted in triplicate form. The seeds were surface sterilized with 0.1% mercuric chloride ( $\text{HgCl}_2$ ) to prevent any contamination. After washing with distilled water they were soaked in 5% bavistin (a systemic fungicide) for 10 – 12 minutes. The selected seeds were placed in 10 cm diameter petridishes lined with filter paper Whatman No. 1 to this 5 ml solution of  $\text{NH}_4(\text{NO}_3)$  at different levels (control, 5 mM) with various concentration of Cd and Hg (control, 10, 20 and 50  $\mu\text{M}$ ) was applied. Distilled water was used as control. The seeds were allowed to germinate for studying various parameters for 10 – 15 days. Three replicates of each treatment were maintained. The chlorophyll and carotenoids were estimated by Arnon's (1949) technique using double beam UV-visible spectrophotometer (Systronics). Proline estimation was carried out by Bates *et al.*, (1973) method, transmittance was read at 520 nm by using double beam UV-visible spectrophotometer (Systronics) and the standard curve was prepared by using pure proline (BDH). Protein estimation was carried out by Folin and Lowry (1975) method, transmittance was read at 750 nm by

using double beam UV-visible spectrophotometer (Systronics) and the standard curve was prepared by using BSA.

### III. RESULT AND DISCUSSION

The effect of Hg was more pronounced than Cd and all concentration and among different types of chlorophyll, chlorophyll 'b' was more effected than chlorophyll 'a'. Data presented in Table 1 indicate that chlorophyll 'a' was reduced upto 0.320 and 0.343  $\text{mg g}^{-1}$  Fw at 10 $\mu\text{M}$  concentration of Hg and Cd respectively as compared to control (0.354  $\text{mg g}^{-1}$  Fw). Chlorophyll 'b' was reduced upto 0.330 and 0.376  $\text{mg g}^{-1}$  Fw at 10 $\mu\text{M}$  concentration of Hg and Cd treatment as compared to control (0.421  $\text{mg g}^{-1}$  Fw). Similarly at 20 $\mu\text{M}$  concentration chlorophyll 'a' reduced upto 0.288 and 0.330  $\text{mg g}^{-1}$  Fw for Hg and Cd as compared to control (0.354  $\text{mg g}^{-1}$  Fw), while chlorophyll 'b' was reduced upto 0.311 and 0.347%  $\text{mg g}^{-1}$  Fw as compared to 0.421  $\text{mg g}^{-1}$  Fw (control) under Hg and Cd treatment respectively and at 50 $\mu\text{M}$  concentration chlorophyll a was reduced upto 0.252 and 0.290  $\text{mg g}^{-1}$  Fw as compared to control (0.354  $\text{mg g}^{-1}$  Fw), while chlorophyll 'b' was reduced upto 0.290 and 0.309  $\text{mg g}^{-1}$  Fw as compared to control (0.421  $\text{mg m}^{-1}$  Fw). The application of nitrogen (Ammonium nitrate) in the nutrient medium proved beneficial for all pigments. Increase in chlorophyll 'b' was more than the chlorophyll 'a'. Carotenoids appears to be more tolerant to heavy metals as compared to other pigments thus showed an increase in the presence of the nitrogen. Thus chlorophyll a, b, and total chlorophyll were drastically reduced under both metal treatments especially at higher level. Similar results have been obtained by Xin Chen *et al.*, 2011 working under Cd stress on pakchoi and mustard plants. Tantrey and Agnihotri (2010) found total chlorophyll reduced considerably due to heavy metal Cd and Hg on Gram (*Cicer arietinum*). Shekar *et al.* (2011) have shown a reduction in chlorophyll content in tomato plants when treated with mercury. With the application of nitrogen carotenoids showed about 4% and 3% increase as compared to control at 50  $\mu\text{M}$  concentration of Cd and Hg. From the data presented in Table 1 it is clear that after the supply of nitrogen all the chlorophyll and carotenoid contents increased as compared to control. Thus in agreement with earlier reporter (Sun *et al.*, 2008, Vajpayee *et al.*, 2000; Mobin and Khan, 2007). Similar results have been obtained in other laboratory studies (Jiang *et al.*, 2007). In this study we find that chlorophyll 'a' content exceeded that of chlorophyll 'b' in all treated plants, which has been proven by other researchers (Mobin and Khan, 2007; Singh *et al.*, 2012). Moreover, the application of heavy metals shows that there occurs a decline in the carotenoid contents in all treated plants and has been proven by various workers (Thapar *et al.*, 2008; Da-Lin *et al.*, 2011; Xin-Chen *et al.*, 2011; Singh *et al.*, 2012). Proline, an amino-acid is well known to get accumulated in wide variety of organisms ranging from bacteria to higher plants on exposure to abiotic stress (Saradhi *et al.*, 1993). A kind of amino acid which could play a therapeutic role in plants. Fennal Plants shows an increase of upto 0.207, 0.268 and 0.468  $\text{mg g}^{-1}$  proline as compared to control (0.182  $\text{mg g}^{-1}$  proline) when treated with the 10 $\mu\text{M}$ , 20 $\mu\text{M}$  and 50 $\mu\text{M}$  concentration of Cd. When treated with Hg, the proline concentration was further enhanced and it was 0.221, 0.282 and 0.498  $\text{mg g}^{-1}$  proline when treated with 10, 20 and

50 $\mu\text{M}$  of Hg as compared to control (0.182  $\text{mg g}^{-1}$ ) proline. However with the application of nitrogen the proline content was reduced to some extent. At higher concentration of Cd and Hg (50 $\mu\text{M}$ ) proline was reduced upto 64% and 65% over control under the application of nitrogen. Similar results were reported by (John *et al.*, 2009; Singh *et al.*, 2012). Heavy metals are known to reduce the protein content of the plants. Data presented in the Table2 demonstrated the deleterious effects of Cd and Hg on protein content. This detrimental effect was more due to Hg as compared to the Cd. As evident from the Table 2 control plants exhibit maximum 0.60  $\text{mg/g}$ , protein content and the minimum 0.42  $\text{mg/g}$  and 0.39  $\text{mg/g}$ , reported in 50 $\mu\text{M}$  Cd and Hg concentration. Protein content was reduced upto 0.55  $\text{mg/g}$  under 10 $\mu\text{M}$  concentration of Cd, 0.50  $\text{mg/g}$  under 20 $\mu\text{M}$  concentration of Cd. Hg proved more harmful for protein content, and reduced it by 0.52  $\text{mg/g}$  under 10 $\mu\text{M}$  Hg, 0.48  $\text{mg/g}$  under 20 $\mu\text{M}$  of Hg. In contrary to Cd and Hg, inclusion of nitrogen ( $\text{NH}_4\text{NO}_3$ ) 0.5mM in nutrient medium increases protein content and proved to be beneficial for plants. Plants showed 24% increases in protein content at 50 $\mu\text{M}$  concentration of Cd and 38% increase at 50 $\mu\text{M}$  of Hg when treated with nitrogen. Our findings are similar to the findings of various workers that there occurs reduction in the protein content by the application of heavy metals ( Balestrasse *et al.*, 2003; Bavi *et al.*, 2011). The reduction in the amount of protein could be due to decrease in protein synthesis or an increase in protein degradation (Balestrasse *et al.*, 2003).

### IV. CONCLUSION

In the present study, exposure of heavy metals (Cd and Hg) to fennel plants affected different parameters like chlorophyll, carotenoids, proline and protein content of this plant. Exposure of metals to the seedlings decreased the total chlorophyll content, carotenoids and proteins contents as the metal concentration goes on increasing. The present results shows that Cd and Hg toxicity increases the proline content of the fennel seedlings with the imposition of low concentration of Cd and Hg (10 $\mu\text{m}$ ) less amount of proline was increased and as the plants are treated with higher doses (20 and 50  $\mu\text{m}$ ) of Cd and Hg there seems to be more increase in the accumulation of proline. In contract to other parameters, in this case the additional supply of nitrogen reduces proline content of the plant.

### REFERENCES

- [1] Aliu, S., Gashi, B., Rusinovei, I. Fetahu, S. and Vataj, R. (2013). Effect of some heavy metals in some morpho-physiological parameters of maize seedlings. American Journal of Biochemistry and Biotechnology, 49(1): 27 – 33.
- [2] Balestrasse, K.B., Benavides, M.P., Gallego, S.M. and Tomaro, M.L. (2003). Effect of cadmium stress on nitrogen metabolism in nodules and roots of soyabean plants. Functional plant biology. 30: 57-64.
- [3] Bavi, K., Kholdebarian, B. and Moradshahi, A. (2011). Effect of cadmium on growth, protein content and peroxidase activity in Pea plants. Pakistan Journal of Botany. 43 (3): 1467-1470.
- [4] Da-lin, L., Kai-qi, H., Jing-jing, M., Wei-wei, Q., Xiu-ping, and Shu-pan, Z. (2011). Effect of cadmium on the growth and physiological characteristics of sorghum plants. African Journal of Biotechnology. 10(70): 15770-15776.
- [5] Husain, A., Z. Baroon, S. Al-Khalafawi, T. Ali-Ati and W. Sawaya. (1995). Heavy metals in fruits and vegetables grown in Kuwait during the oil well fires. Arab Gulf Journal of Science Research. 13: (3) 535-542.

- [6] John, R., Ahmad, P., Gadgil, K. and Sharma, S. (2009). Effect of cadmium and lead on growth, biochemical parameters and uptake in *Lemna polysshiza* L. *Plant Soil and Environment*. 54: 262-270.
- [7] Kaur, G.J. and Arora, D.S. (2010). Antibacterial and phytochemical screening of *Anethum graveolens*, *Foeniculum vulgare* and *Trachyspermum ammi*. *BMC complementary and Alternative Medicine*: 9-30.
- [8] Mobin, M. and Khan, N.A. (2007). Photosynthetic activity, pigment composition and antioxidative response of two mustard (*Brassica juncea*) cultivars differing in photosynthetic capacity subjected to cadmium stress. *Journal of Plant Physiology*. 164: 601-610.
- [9] Prajapati, N.D., Prajapati, T. and Jaipura, S. (2005). *Advances in medicinal plants*. Asian Medicinal Plants and Health Care Trust Publishers. 1: 222.
- [10] Saradhi, P.P. and Alia, V.B. (1993). Inhibition of mitochondrial electron transport is the prime cause behind proline accumulation during mineral deficiency in *Oryza sativa*. *Plant and Soil*. 155/156: 465-468.
- [11] Shekar, C.C., Sammaiah, D., Shastree, T. and Reddy, K.J. (2011). Effect of mercury on Tomato growth and yield attributes. *International Journal of Pharma and Bioscience*. 2 (2): 385-364.
- [12] Singh, G., Agnihotri, R.K., Sharma, R. and Ahmad, M. (2012). Effect of lead and nickel toxicity on chlorophyll and proline content of urd (*Vigna mungo* L.) seedlings. *International Journal of Plant Physiology and Biochemistry*. 10 (10): 30: 32.
- [13] Sinha, S. Gupta, M. Chandra, P. (1996). Bioaccumulation and biochemical effect of mercury in the plant of *Bacopa monnieri* L. *Environ Toxicol water Qurl*. 11 : 105 – 112.
- [14] Sun, Y., Zhou, Q. and Chunyan, D. (2008). Effect of cadmium and arsenic on growth and metal accumulation of Cd-hyperaccumulator *Solanum nigrum* L. *Bioresource Technology*. 99: 1103-1110.
- [15] Tantrey, M.S. and Agnihotri, R.K. (2010). Chlorophyll and proline content of gram (*Cicer arietinum* L.) under cadmium and mercury treatment. *Research Journal of Agricultural Sciences*. 1 (2): 119-122.
- [16] Tantrey, M.S. and Agnihotri, R.K. (2012). Reduction in germination and seedling growth of *Cicer arietinum* L. caused by mercury and cadmium treatments. *Flora and Fauna*. 18 : 2.
- [17] Thapar, R., Srivastava, A.K., Bhargava, P., Mishra, Y. and Rai, L.C. (2008). Impact of different abiotic stress on growth, photosynthetic electron transport chain, nutrient uptake and enzyme activities of Cu-acclimated *Anabaena doliolum*. *Journal of Plant Physiology*. 165: 306-316.
- [18] Vajpayee, P., Tripathi, R.D., Rai, U.N., Ali, M.B. and Singh, S.N. (2000). Chromium (VI) accumulation reduces chlorophyll biosynthesis, nitrate reductase activity and protein content in *Nymphaea alba* L. *Chemosphere*. 41: 1075-1082.
- [19] Wang Hong-Wei; Shi Ya-Qi; Huang Feng and Yuan Lin (2008). Effect of HgCl<sub>2</sub> short time treatment on photosynthesis in broad bean leaves. *Acta Agronomica Sinica*. 34 (1): 157 – 162.
- [20] Xin Chen, Wang, J., Shi, Y., Zhao, M.Q. and Chi, G.Y. (2011). Effect of Cd on growth and photosynthetic activities in pakchoi and mustard. *Botanical Studies*. 52: 41-46.

#### AUTHORS

**First Author** – Mohsin Anayat, Department of Botany, School of Life Sciences, Khandari, Agra, Dr. B.R. Ambedkar University Agra-282002

**Second Author** – Rajneesh Kumar Agnihotri, Department of Botany, School of Life Sciences, Khandari, Agra, Dr. B.R. Ambedkar University Agra-282002, Email id : rk\_agnihotri@rediffmail.com

**Third Author** – Rashmi Vamil, Department of Botany, School of Life Sciences, Khandari, Agra, Dr. B.R. Ambedkar University Agra-282002

**Fourth Author** – Sanjay Kumar, Department of Botany, School of Life Sciences, Khandari, Agra, Dr. B.R. Ambedkar University Agra-282002

**Fifth Author** – Rajendra Sharma, Department of Botany, School of Life Sciences, Khandari, Agra, Dr. B.R. Ambedkar University Agra-282002

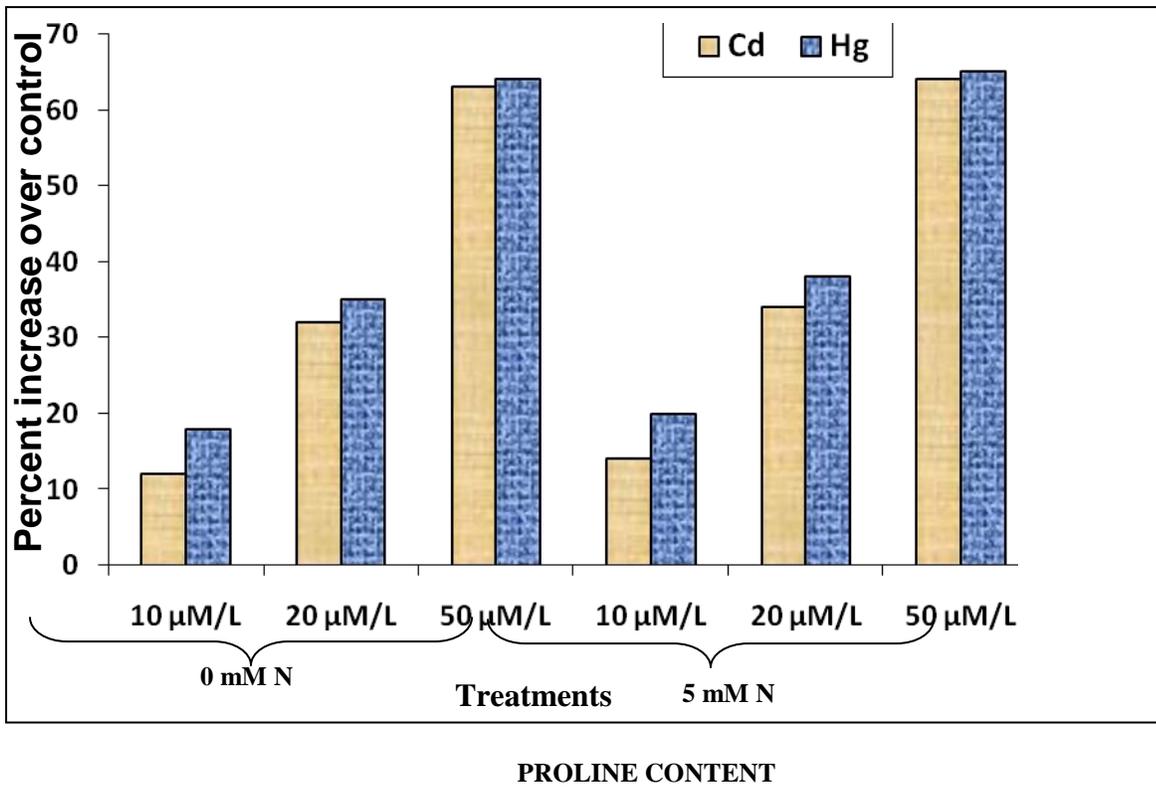


Fig 1. : Effect of heavy metals and nitrogen interaction on proline content ( $\text{mg g}^{-1}$  FW) in *Foeniculum vulgare L.*

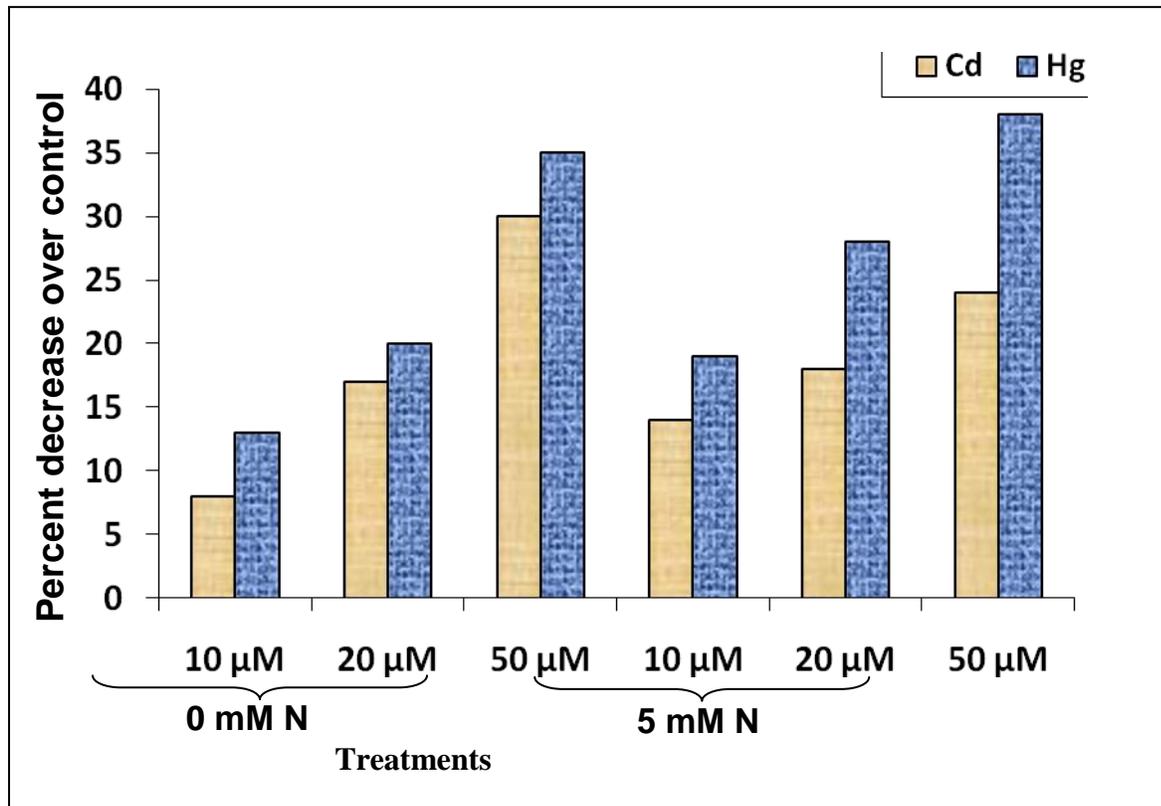


Fig. 2 : Effect of heavy metals interaction on protein content in *Foeniculum vulgare* L. with and without nitrogen

Table 1 : Effect of Cadmium and mercury on pigment composition ( $\text{mg g}^{-1}$  FW) in Fennel (*Foeniculum vulgare* L.) grown with or without nitrogen.

Metals	Concentration ( $\mu\text{M}$ )	Chlorophyll a		Chlorophyll b		Total Chlorophyll		Carotenoids	
		0 mM N	5 mM N	0 mM N	5 mM N	0 mM N	5 mM N	0 mM N	5 mM N
Control	0	0.354	0.402	0.421	0.483	0.775	0.885	1.547	1.646
Cd	10 $\mu\text{M}$	0.343	0.389	0.376	0.465	0.719	0.854	1.520	1.603
	20 $\mu\text{M}$	0.330	0.370	0.347	0.412	0.677	0.782	1.492	1.562
	50 $\mu\text{M}$	0.290	0.350	0.309	0.391	0.599	0.741	1.418	1.497
Hg	10 $\mu\text{M}$	0.320	0.380	0.330	0.415	0.650	0.795	1.485	1.579
	20 $\mu\text{M}$	0.288	0.337	0.311	0.400	0.599	0.737	1.439	1.490
	50 $\mu\text{M}$	0.252	0.310	0.290	0.363	0.542	0.673	1.376	1.421

N – Nitrogen

Data are average of 3 replicates.

**Table 2 : Effect of Cadmium and mercury on proline content (mg g<sup>-1</sup> FW) and protein content in Fennel (*Foeniculum vulgare* L.) in shoots grown with or without nitrogen.**

Metals	Concentration (µM)	Proline content (mg g <sup>-1</sup> FW)		Protein content (mg g <sup>-1</sup> FW)	
		0 mM N	0 mM N	5 mM N	5 mM N
Control	0	0.182	0.60	0.72	0.165
Cd	10 µM	0.207	0.55	0.62	0.192
	20 µM	0.268	0.50	0.59	0.250
	50 µM	0.486	0.42	0.55	0.462
Hg	10 µM	0.221	0.52	0.58	0.206
	20 µM	0.282	0.48	0.52	0.265
	50 µM	0.498	0.39	0.45	0.477

N – Nitrogen

Data are average of 3 replicates.