

Almond leaves for the one-pot Biofabrication of silver nanoparticles: Characterization and larvicidal application

Ayandiran D. Aina^{a*}, Oluwafayoke Owolo^{a,d}, Morenike Adeoye-Isijola^a, Folasade O. Aina^b, Omeiza Favour^d and Aderiike, G. Adewumi^f

^aDepartment of Microbiology, Babcock University, PMB 21244, Ikeja, Lagos, Nigeria

^bDepartment of Maternal and Child Health, Babcock University, PMB 21244, Ikeja, Lagos, Nigeria

^cDepartment of Pure and Applied Biology, Ladoko Akintola University of Technology, PMB 4000, Ogbomoso, Nigeria

^dDepartment of Pharmaceutical Microbiology, University of Ibadan, PMB 5116, Ibadan, Oyo State, Nigeria

^fDepartment of Basic and Applied Sciences, Babcock University, PMB 21244, Ikeja, Lagos, Nigeria

Corresponding author: Ayandiran, D. Aina

Telephone number: +2348033565433

Email address: ainaa@babcock.edu.ng

DOI: 10.29322/IJSRP.8.11.2018.p8378

<http://dx.doi.org/10.29322/IJSRP.8.11.2018.p8378>

Abstract

This work reports the synthesis of silver nanoparticles (AgNPs) using the leaf extract of Almond plant. Larvicidal activities of the biosynthesized AgNPs were evaluated against the fourth instar larvae of mosquito *Aedes aegypti*. The biosynthesized AgNPs were characterised using UV-Visible, Fourier Transmission Infrared Spectroscopy (FTIR), Energy Dispersive X-ray Spectroscopy (EDX) and Field Emission Scanning Electron Microscope (FESEM). The absorption spectrum of the synthesized AgNPs showed a maximum spectrum of 430 nm while FTIR analysis showed different functional groups present on the surface of the AgNPs with broad peak between 3000 and 3800 cm^{-1} . The FESEM showed a large number of spherically shaped nanoparticles with sizes ranging from 8.34 to 78.96 nm. The 50% (LC_{50}) lethal concentration of the nanoparticles after 12 h was 13.54 $\mu\text{g/ml}$. This study showed that almond leaf can be used for synthesis of AgNPs thus adding to the available drugs that could be used in combating multidrug resistant pathogens.

Key words: Almond leaves, larvicidal, silver nanoparticles, green synthesis

INTRODUCTION

Nanotechnology is one of the main logical fields today since it combines learning from the fields of Physics, Chemistry, Biology, Medicine, Informatics, and Engineering. It is a rising technological field with remarkable potential to lead in great breakthrough that can be connected, all things considered. Novel nano and biomaterials, and nano gadgets are created and controlled by nanotechnology instruments and systems, which research and tune the properties, reactions, and elements of living and non-living matter, at sizes beneath 100 nm [1].

The term nanotechnology originates from the blend of two words: the Greek numerical prefix nano alluding to a billionth and the word technology. Thus, Nanotechnology or Nanoscaled Technology is for the most part thought to be at a size below 0.1 μm or 100 nm (a nanometer is one billionth of a meter, 10^9m). Nanoscience studies the properties and reactions of materials at nuclear, atomic, and macromolecular scales, and generally at sizes in the vicinity of 1 and 100 nm. In this scale, and particularly beneath 5

nm, the properties of matter vary essentially (i.e., quantum-scale impacts assume a critical part) from that at a greater particulate scale. Nanotechnology is then the plan, the control, the building, the generation and application of structures, gadgets and frameworks of about or less than 100 nm [2-3].

Nanoparticles have been reported as having various medicinal applications. Studies such as Nasrollahi *et al.* [4] have shown the antifungal activities of silver nanoparticle. In another recent paper, Adelere *et al.* [5] reported the antimicrobial activity of a green synthesized nanoparticle. In addition, nanoparticles have been reported to have anti-inflammatory [6], antiangiogenesis [7], antiviral [8] and antiplatelet activity [9]. Plant extracts have recently been used for nanoparticles green synthesis since they are rich in bioactive compounds [5, 10-12] and hence this study was also aimed at the synthesis and application of nanoparticles gotten from plant extract.

The leaf of the almond plant is a potential wellspring of bioactives; it is intriguing to assess the viability of the aqueous extraction from almond leaves (frequently disposed of as an agrowaste) towards the biogenic synthesis of silver nanoparticle. Almond (*Prunus dulcis* L.), a specie of *Prunus*, belongs to the subfamily Prunoideae of the family Rosaceae. Nutritiously and therapeutically, almond has been accounted for to be a valuable food item [13-14].

As a result of lacking information from works on the effect of *P. dulcis* on larvicidal activity and in addition to the very little utilization of this plant in the synthesis of nanoparticles, the principal aim of this investigation is, hence, to use the aqueous extract of the plant leaves for synthesis of silver NPs and to assess the in vitro larvicidal activity of the biosynthesized silver nanoparticle.

MATERIAL AND METHODS

Sample collection

Fresh *Prunus dulcis* leaves were collected from the local area of the University campus. It was washed thoroughly with distilled water several times to remove dust and dried under shade ($30^{\circ}\text{C} \pm 2^{\circ}\text{C}$) for 5 days. The dried leaves were cut into small pieces, and blended into powder. The powdered samples were kept in air tight containers at room temperature for further use.

Collection and Identification of Mosquito Larva

The mosquito larvae were gotten from a stagnant drainage on the University campus and were kept in a closed container for safety precautions in case of the fast emergence of the larvae into an adult mosquito. The target mosquito larva in this study was the fourth instar larva of dengue carrying mosquito *Aedes aegypti*. Identified *Aedes aegypti* mosquito larvae (Identification was done by a medical entomologist) were separated from the other mosquito species and were placed in a water- filled plastic moulder.

Preparation of extract

One gram of the milled leaves of *Prunus dulcis* was weighed and suspended in 100ml of distilled water. The extract was obtained by heating in water bath at 60°C for 1 hour. The extract was filtered using Whatman No. 1 filter paper and then centrifuged at 4000RPM for 20minutes. The supernatants were collected and used without further purification [15]

Synthesis of Silver nanoparticles

1mM aqueous solution of silver nitrate was prepared for synthesis of silver nanoparticles. Approximately 1 ml of the extract was added to a reaction vessel containing 40ml of a 1mM silver nitrate (AgNO_3) solution to reduce the amount of silver

ions. The reaction was carried out under static condition at room temperature ($30 \pm 2^\circ\text{C}$) for 2hr. The formation of AgNPs was observed as a change in the solution colour [15]

Characterization of Synthesized Silver nanoparticles

The formation of the synthesized nanoparticles was confirmed by measuring its absorbance spectrum using UV–Visible spectrophotometer (Cecil, USA) operated at 190–1100 nm. The identity of the biomolecules that took part in the green synthesis was determined by FTIR spectroscopy. The measurements were performed between 4000–400 cm^{-1} to see the attachment of biomolecules on the surface of the AgNPs using Shimadzu FTIR spectrometer, model 8400S (Shimadzu, Japan). To achieve this, purified silver nanoparticles were dried and blended with KBr in the ratio 5: 95 to form a pellet which was used for the measurement. The size, morphology, and elemental composition of the synthesized nanoparticles were unravelled by Field Emission Scanning Electron Microscopy (FESEM) and EDX analyses. The Field Emission Scanning Electron Microscopy (FESEM) micrograph was obtained as follows. A drop of nanoparticles in suspension was placed on a 200 mesh hexagonal copper grid (3.05 mm) (Agar Scientific, Essex, UK) coated with 0.3 % formvar dissolved in chloroform. The particles were allowed to settle for 3–5 min on the grid, the excess liquid flicked off with a wick of filter paper and the grids were then air dried before FESEM viewing. Micrograph was obtained using a JEM-1400 (JEOL, USA) operating at 200 kV.

Larvicidal Activity of the synthesized Nanoparticle

The efficacy of the synthesized nanoparticle as larvicide against the dengue-vector *Aedes aegypti* mosquito was evaluated in a dose-response bioassay against the fourth instar larvae in accordance with the guidelines of World Health Organization [16]. The larvicidal activity was conducted in triplicate by exposing five *Aedes aegypti* mosquito larvae to 300 μl of each of the graded concentrations (20, 40, 60, 80 and 100) of Ag-NPs at room temperature ($30 \pm 2^\circ\text{C}$). In the control experiment, the larvae were exposed to sterile distilled water under the same conditions. The number of death was plotted against concentration of the Ag-NPs. The effects of the synthesized nanoparticles were monitored through carefully counting the number of dead larvae after 12 hours of treatment, and the percentage mortality was computed. Probit Analysis was used to calculate LC_{50} value to determine lethal concentrations of the synthesized nanoparticles on *Aedes aegypti* mosquito larvae after 12 hours of treatment.

RESULTS AND DISCUSSION

Synthesis and Characterization of the Silver nanoparticle

The phytosynthesis of the silver nanoparticles was catalyzed by the aqueous extract of the almond leaves. Within the first 10 mins, a colour change was observed. The initial solution which was colorless was transformed to light brown and then stabilized at a dark brown color as indicated in figure 1. It is known that when the surface plasmon vibrations in silver nanoparticles are excited, the silver nanoparticles exhibit some yellowish brown color in the aqueous solution [17].

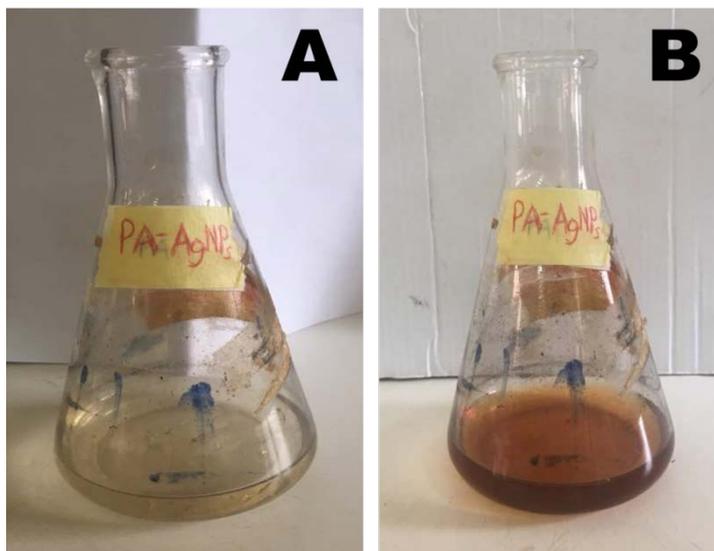


Figure 1: Synthesis of the Almond leaf extract mediated silver nanoparticles (a) immediately after the addition of the almond leaf extract to the silver nitrate; (b) Formation of deep brown colouration after 30 min.

The biosynthesized silver nanoparticle showed a maximum absorbance wavelength at 430nm which is indicated in figure 2, a value within the range previously reported for AgNPs [17]

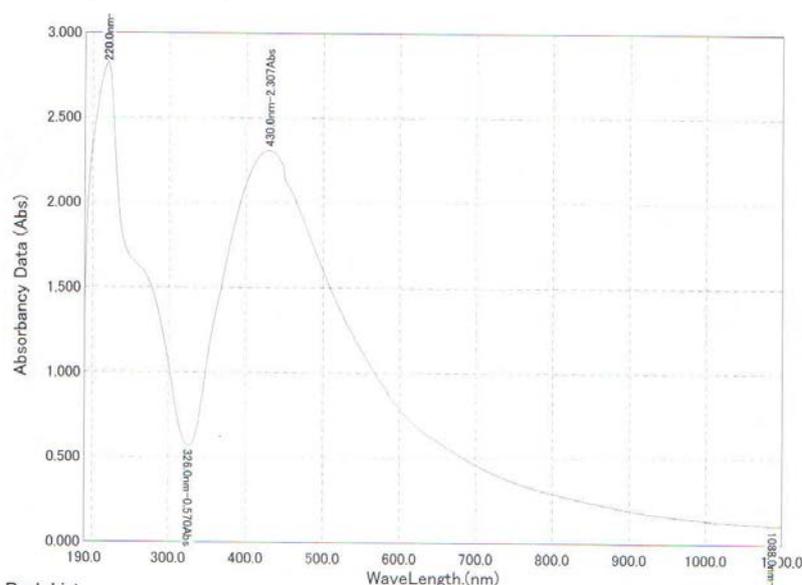


Figure 2: UV-Vis spectra of silver nanoparticles synthesized from almond leaf extract

The NP shows a broad peak between 3000 and 3800 cm^{-1} , which are identified as those of O-H vibrations and/or N-H stretching associated with N-substituted amide [18], 2359 cm^{-1} peak is that of CO_2 from air, the 2000 cm^{-1} peak is probably from C=N and/or C=O bond, the distinct peak at 1635 cm^{-1} is the -N-H bend of amino acids/proteins [19-20], and the one at 1384 cm^{-1} is due to in plane bending of alkenes and aromatics [21]. The Ag-O stretching modes are observed at 669 cm^{-1} and 420 cm^{-1} [22]. In essence, the FTIR analyses suggested the presence of phytochemicals on the surface of the NPs as capping agents, which are then released systematically as drugs in the study.

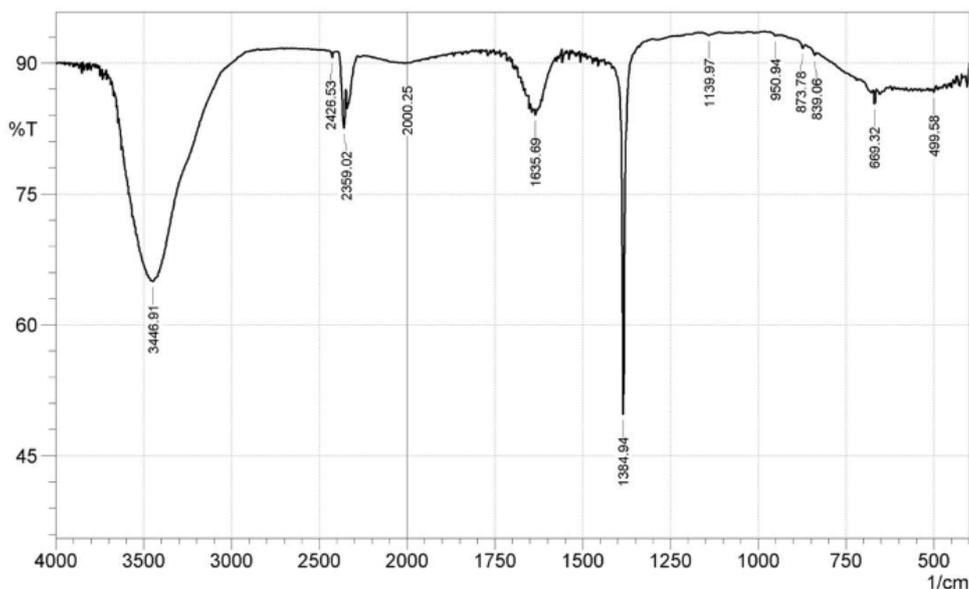


Figure 3: The FTIR spectra of the silver nanoparticles synthesized from almond leaf extract

The chemical analysis of the almond leaf extract - mediated silver nanoparticle was shown (Figure 4) by the EDX spectroscopy. Elemental signals were observed around the Ag atom within the ranges of 2.5 to 3.2keV. Other elements present include copper, oxygen, and carbon). Similar observations have been reported by other researchers [23-24] with silver signals at the range of 1.5 to 5.0 keV

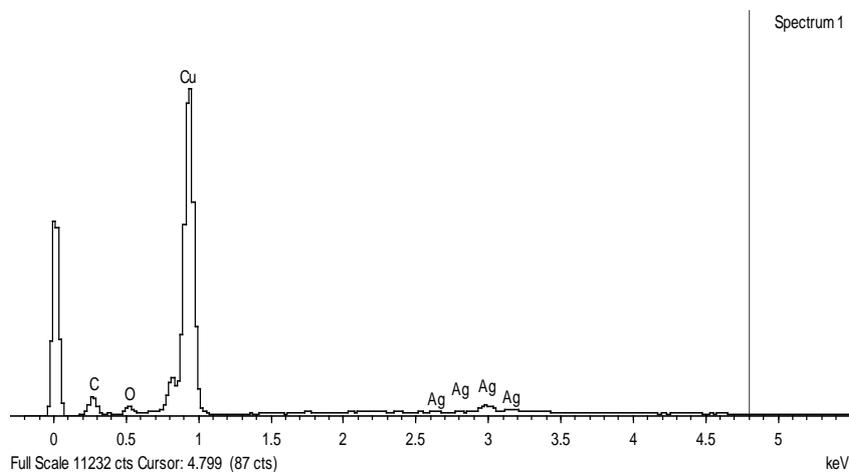


Figure 4: Energy dispersive spectra of the synthesized AgNPs

The size and morphology of the biosynthesized nanoparticle were captured by (Figure 5) FESEM. The particles which ranged from 8.34 to 78.96 nm were spherical in shape. This size range falls within the ranges reported when other plants were used in the biosynthesis of silver nanoparticle [15, 25]

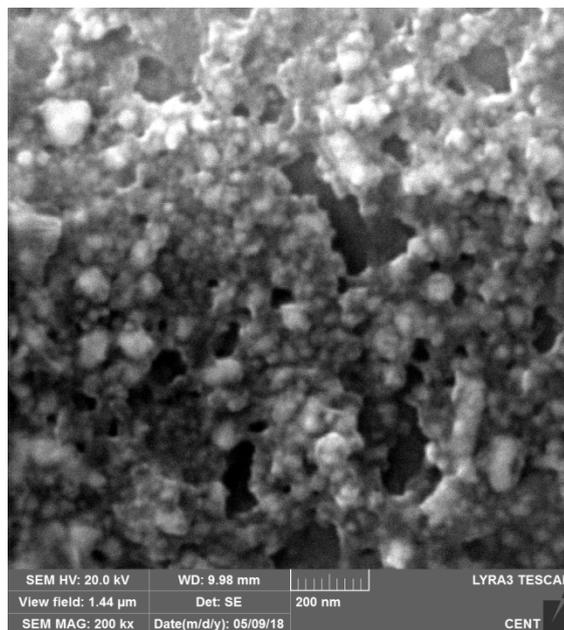


Figure 5: Field Emission Scanning Electron Micrograph of the Synthesized AgNPs

Larvicidal Activity of the synthesized Nanoparticle

The biosynthesized silver nanoparticle was found to be highly toxic to the 4th instar larvae *Aedes aegypti*. After 12 hours of incubation, mortality percentage in the 20 μ g/ml and 40 μ g/ml was found to be 80 and 86.6 respectively while the 60 μ g/ml, 80 μ g/ml and 100 μ g/ml showed 100% mortality. The control group however had no mortality. Furthermore, the silver nanoparticles exhibited a concentration dependent activity against mosquito larvae since the percentage mortality were observed to increase with increasing concentrations of the biosynthesized nanoparticles. The 50% lethal concentration (LC₅₀) of the nanoparticles after 12h was 13.54 μ g/ml. In previous study, Lateef *et al.* [26] also reported similar result when cell free extract of *Bacillus safensis* was used in the synthesis of silver nanoparticles. The resulting nanoparticle was reported to have larvicidal activity. The larvicidal activities of the AgNPs could be due to penetration of the particles to impair cellular metabolism due to their binding to DNA and enzymes. There are increasing evidences to show that metallic nanoparticles that are synthesized through the green route by using plant-based extracts have important roles to play as emerging nanotools in the control of mosquitoes of medical and veterinary importance [27]

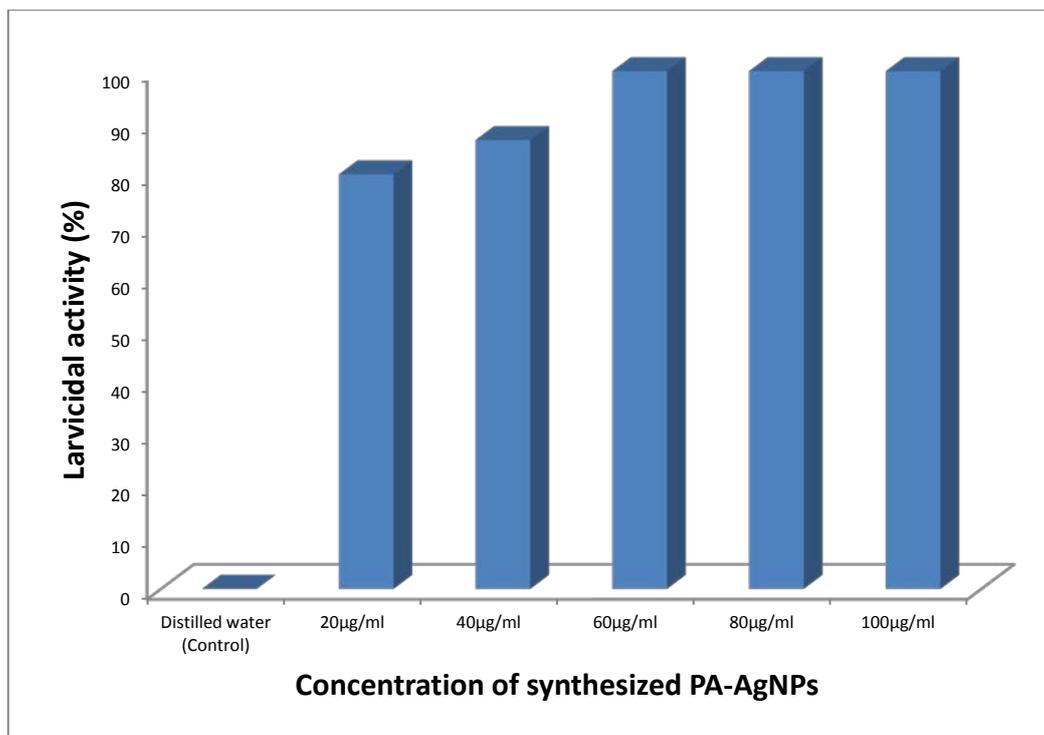


Figure 6: The larvicidal activity of the synthesized AgNPs

CONCLUSION

In this work, almond leaf extract was used to synthesize nanoparticles. The synthesised AgNPs was analysed using UV Spectrophotometer, FTIR, FESEM and EDX. The biosynthesised silver nanoparticle had larvicidal activities against *Aedes aegypti* larvae. Thus almond leaf extract mediated-AgNPs could have various pharmaceutical applications.

ACKNOWLEDGEMENTS

The authors thank the department of Microbiology, Babcock University, Ilisan-Remo, Nigeria for her assistance during this study.

FUNDING

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors

CONFLICT OF INTEREST

The authors declare that they have no conflict of interest.

REFERENCES

1. Logothetidis, S. (2014). Nanostructured Materials and Their Applications. Berlin: Springer Berlin.
2. Sattler, K.D. (2010) Handbook of Nanophysics, Principles and Methods. CRC, New York,
3. Bhushan, B. (2004) Handbook of Nanotechnology. Springer, Berlin: Springer Berlin.
4. Nasrollahi, Y.-K., Kim, B. H., & Jung, G. (2009). Antifungal Activity of Silver Nanoparticles on some fungi. *Plant Disease*, 93(10), 1037–1043. <https://doi.org/10.1094/PDIS-93-10-1037>

5. Adelere, A. I., Lateef, A., Oyeyemi, A. D., Ramatu, A., Usman, N., and David, B. J. (2017). Biosynthesis of silver nanoparticles using aqueous extract of *Buchholzia coriacea* (wonderful kola) seeds and their antimicrobial activities, *Annals Food Science and Technology* 18(4): 671–679.
6. Baharara, J., Asadi-Samani, M., Ramezani, T. and Mousavi, M. (2017). Antioxidant and anti-inflammatory activity of green synthesized silver nanoparticles using *Salvia officinalis* extract. *Annals of Tropical Medicine and Public Health*, 10(5), p.1265.
7. Baharara, J., Namvar, F., Mousavi, M., Ramezani, T. and Mohamad, R. (2014). Anti-Angiogenesis Effect of Biogenic Silver Nanoparticles Synthesized Using *Salvia officinalis* on Chick Chorioalantoic Membrane (CAM). *Molecules*, 19(9), pp.13498-13508.
8. Park, I., Kwon, Y., Ryu, W. and Ahn, B. (2014). Inhibition of hepatitis B virus replication by ligand-mediated activation of RNase L. *Antiviral Research*, 104, pp.118-127.
9. Shrivastava S, Bera T, Singh S, Singh G, Ramachandrarao P, Dash D. (2009) Characterization of Antiplatelet Properties of Silver Nanoparticles. *ACS Nano*. 3, 1357-1364.
10. Ahmed, S., Saifullah, Ahmad, M., Swami, B. L., and Ikram, S. (2016). Green synthesis of silver nanoparticles using *Azadirachta indica* aqueous leaf extract. *Journal of Radiation Research and Applied Sciences* 9(1): 1–7.
11. Thirumurugan, A., Tomy, N. A., Ganesh, R. J., and Gobikrishnan, S. (2010). Biological reduction of silver nanoparticles using plant leaf extracts and its effect on increased antimicrobial activity against clinically isolated organisms. *Der Pharma Chemica* 2(6): 279-284.
12. Benakashani, F., Allafchian, A. R., and Jalali, S. A. H. (2016). Biosynthesis of silver nanoparticles using *Capparis spinosa* L. leaf extract and their antibacterial activity. *Karbala International Journal of Modern Science* 2(4): 251–258.
13. Wijeratne SSK, Abou-Zaid MM, Shahidi F (2006). Antioxidants polyphenols in almond and its coproducts. *Journal of Agriculture and Food Chemistry*, 54, 312–318.
14. Sarwar, S. (2012). Antioxidant characteristics of different solvent extracts from almond (*Prunus dulcis* L.) shell. *Journal of Medicinal Plants Research*, 6(17), 3311–3316. <https://doi.org/10.5897/JMPR11.1723>
15. Lateef A., Folarin B., Oladejo S., Akinola P., Beukes L. & Gueguim-Kana E. (2018). Characterization, antimicrobial, antioxidant, and anticoagulant activities of silver nanoparticles synthesized from *Petiveria alliacea* L. leaf extract. *Preparative Biochemistry and Biotechnology*. <https://doi.org/10.1080/10826068.2018.1479864>
16. WHO. (2008). Guidelines for laboratory and field testing of mosquito larvicides. *World Health Organization*: 1–41.
17. Krishnaraj C., Jagan E. G., Rajasekar S., Selvakumar P., Kalaichelvan P. T., & Mohan N. (2010). Synthesis of silver nanoparticles using *Acalypha indica* leaf extracts and its antibacterial activity against water borne pathogens. *Colloids Surf B Biointerfaces*. 76: 50–56.
18. Unuabonah, E.I., Adie, G.U., Onah, L.O. and Adeyemi, O.G. (2009). Multistage optimization of the adsorption of methylene blue dye onto defatted *Carica papaya* seeds. *Chemical Engineering Journal* 155 (3): 567-579
19. Lin W., Yu D. & Yang M. (2005). pH-sensitive polyelectrolyte complex gel microspheres composed of chitosan/sodium tripolyphosphate/dextran sulfate: swelling kinetics and drug delivery properties. *Colloids Surf B Biointerfaces*. 44: 143-151.
20. Sun L., Du Y., Chen L., Huang R. & Chen X. (2004). The synthesis of carboxymethylchitosan hydrogel and the application in drug controlled release systems. *Acta Polym Sin.* 8: 191–195
21. Rahmaniyan F., Shamel A. & Shafaghatlonbar A. (2014). Evaluation of Biologically Synthesized Silver Nanoparticles by the Bioreduction Method. *Synth React Inorg M.* 45: 1495-1500.
22. Taghavi Fardood S., Ramazani A., Moradi S. & Azimzadeh Asiabi P. (2017). Green synthesis of zinc oxide nanoparticles using arabic gum and photocatalytic degradation of direct blue 129 dye under visible light. *J Mater Sci - Mater El.* 28: 13596-13601.
23. Islam N., Amin R., Shahid M., Amin M., Zaib S. & Iqbal J. (2017). A multi-target therapeutic potential of *Prunus domestica* gum stabilized nanoparticles exhibited prospective anticancer, antibacterial, urease-inhibition, anti-inflammatory and analgesic properties. *BMC Complement Altern Med.* 276: 1-17.

24. Puchalski P., Dabrowski P., Olejnikzac W., Krukowski P., Polanski K., & Kowalczyk K. (2007). The study of silver nanoparticles by scanning electron microscopy, energy dispersive X-ray analysis and scanning tunnelling microscopy. *J. Mater. Sci.* 25: 23-31.
25. Kumar C., Yugandhar P. & Savithamma N. (2016). Biological synthesis of silver nanoparticles from *Adansonia digitata* L. fruit pulp extract, characterization, and its antimicrobial properties. *J Intercult Ethnopharmacol.* 5: 79-85.
26. Lateef, A., Ojo, S. A., Akinwale, A. S., Azeez, L., Gueguim-Kana, E. B., and Beukes, L. S. (2015). Biogenic synthesis of silver nanoparticles using cell-free extract of *Bacillus safensis* LAU 13: Antimicrobial, free radical scavenging and larvicidal activities. *Biologia (Poland)* 70(10): 1295–1306.
27. Benelli, G. (2015). Plant-mediated biosynthesis of nanoparticles as an emerging tool against mosquitoes of medical and veterinary importance: a review. *Parasitology Research*, 115(1), pp.23-34.