

Speciation of *Candida* Species Isolated From Clinical Specimens by Using Chrom Agar and Conventional Methods

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Abstract- *Candida* spp especially non albicans *Candida* are increasingly being isolated from clinical specimens. The conventional methods of identification are time consuming and difficult to perform. The study was done to evaluate the performance of conventional identification method (phenotypic and biochemical) and commercially available chromogenic *Candida* speciation media (CHROM agar) for the identification of medically important yeast and yeast-like organisms in a routine clinical microbiology laboratory. A total of 60 yeast strains were included during the one and half years study period. The conventional methods used for speciation of yeast isolates were germ tube test, chlamyospore formation test on corn meal agar, sugar fermentation test and sugar assimilation test and were compared against chromogenic agar medium (CHROM agar). *Candida albicans* (51.6%) was the most common *Candida* species, followed by *C. tropicalis* (25%), *C. krusei* (16.6%) and *C. glabrata* (6.6%). Agreement between the conventional method and chromogenic methods was 96% for *C. albicans*, 100% for *C. tropicalis*, 100% for *C. krusei* and 100% *C. glabrata*. *C. albicans* was the most common single species isolated. However, species other than *C. albicans* are gaining clinical significance (48% of all isolates in the present study). CHROM agar is a convenient and rapid method of identification of *Candida* species even in resource poor settings.

Index Terms- *Candida*, CHROM agar, identification, sugar assimilation test

I. INTRODUCTION

Candida spp. are the members of the normal flora of the skin, mucous membranes, and gastrointestinal tract. They are endogenous opportunists which cause secondary infection in individuals with some underlying immunocompromised conditions. Candidiasis is a common fungal disease found in humans affecting mucosa, skin, nails and internal organs of the body.

Candida albicans is generally considered the major pathogen among the *Candida* species. An increase in the prevalence of non-albicans species has been noted during the last decades.^{1,2} There is growing evidence of the increasing use of azoles causing this epidemiological shift. Characterization to species level helps to identify those strains which might be intrinsically resistant to some of the antifungal agents.^{3,4} Speciation of *Candida* isolates is conventionally done by germ tube test, sugar assimilation and sugar fermentation tests. Newer methods include CHROM agar,

API systems, Vitek 2 ID system and molecular methods.^{5,6,7} Germ tube test is a rapid method to differentiate *C. albicans* and *C. dubliniensis* from other *Candida* spp. For further speciation chlamyospore formation test, sugar fermentation test and sugar assimilation test can be done. But these tests are time consuming and labour intensive. Among the newer tests, CHROM agar is rapid and cost effective as compared to other expensive systems like API systems, Vitek 2 ID system and molecular methods.

In the present study, we speciated *Candida* isolates using germ tube test, chlamyospore formation test, sugar fermentation test, sugar assimilation test and also evaluated the performance of commercially available chromogenic *Candida* speciation media i.e, CHROM agar.

II. MATERIALS AND METHODS

The present study was carried out between October 2011 and April 2013 in the Department of Microbiology, SGT Medical College, Budhera, Gurgaon. A total of 60 consecutive *Candida* isolates from various clinical specimens like high vaginal swab, urine, blood, sputum, pus, catheter tip, ear swab from patients with candidiasis and stool sample from patients with antibiotic associated diarrhoea were included in the study. These specimens were processed for the isolation of *Candida* spp. using standard Mycology methods.⁵ Gram staining was performed from direct specimen and the specimens were inoculated on Sabouraud's dextrose agar slants, incubated at 37°C for 24 hrs. Germ tube test was done and the positives identified were either *C. albicans* or *C. dubliniensis*. *C. albicans* were further identified by growth at 45°C and Chlamyospore formation on corn meal agar.⁷ All the 60 isolates were subjected to Sugar fermentation test and Sugar assimilation test for final confirmation of species.

Simultaneously the *Candida* spp. were inoculated on CHROM agar and incubated at 37°C for 24 hrs and the species were identified by type and colour of the colonies on CHROM agar media as per manufacturer's instructions. (Table 1.)

III. RESULT

A total of 60 *Candida* spp. were isolated from various clinical specimens. Table 2 gives the distribution and sources of *Candida* spp. identified by the gold standard conventional method. *Candida albicans* (52%) was the most common species isolated. Among the non albicans candida, *C. tropicalis* (25%) was the commonest followed by *C. krusei* (17%) and *C. glabrata* (7%). (Table 2).

These 60 strains were also subjected to identification using CHROM agar. The results along with sensitivity and specificity of Chrome agar for various species are given in table 3. There was an agreement in identification by CHROM agar method in 59 (98%) strains. Only one strain which was identified as *C. glabrata* by the sugar assimilation test was identified as *C. albicans* by chrome agar method. The sensitivity and specificity of CHROM agar was 100% for *C. Tropicalis* and *C. Krusei*, for *C. albicans* the sensitivity was 100% and specificity was 96% and for *C. glabrata* the sensitivity was 75% and specificity was 100%. (Table 3).

IV. DISCUSSION

The potential clinical importance of species-level identification has been recognized as *Candida* species differ in the expression of virulence factors and antifungal susceptibility. Non albicans candida are on the rise due to increasing immunocompromised states. Non albicans *Candida* are more resistant to fluconazole, therefore species level identification has a direct impact on choice of empirical antifungal treatment. Also there may be geographic variation in the species isolated which necessitates that we have data on the distribution of candida species in different geographic regions. In the present study *C. albicans* predominated i.e., 52%. Predominance of *C. albicans* was also seen in a study.⁸ However higher incidence of non albicans candida ranging from 54 - 74% have been seen in various studies.^{4,9,10} Among the non albicans species, *Candida tropicalis* is reported to be the most predominant species as discussed elsewhere. In our study also *C. tropicalis* was the most common non albicans species.

For differentiation between different species of candida conventionally Germ tube test, chlamydospore formation, sugar fermentation and assimilation tests are being used which are laborious and time consuming. CHROM AGAR is a rapid method to differentiate between different candida species. It facilitates the detection and identification of candida species from mixed culture and provides result in 24-48 hours. In our study, sensitivity and specificity of CHROM agar for *Candida albicans* were 100% and 96%, *C. tropicalis* were 100% and 100%, *C. krusei* were 100% and 100% and *C. glabrata* 75% and 100% respectively. A sensitivity of 80% for *C. tropicalis*, and 89% for *C. albicans* has been reported in a study who also reported that misidentification of *C. tropicalis* was seen due to difficulty in interpretation of green color.⁶ We, however, did not face any such difficulty and our results were consistent with the results of conventional methods. CHROM agar has the advantage of being technically simple, rapid and cost effective as compared to the conventional methods.

Being a rural hospital and medical college, our study had its own limitations of small sample size, inability to perform antifungal susceptibility tests. However CHROM agar has proved to be a valuable method for identification of *Candida* species even in resource poor settings.

V. CONCLUSION

Along with *Candida albicans*, non albicans *Candida* spp like *C. tropicalis*, *C. krusei* and *C. glabrata* are increasingly being isolated from clinical specimens. CHROM agar is a simple, rapid and inexpensive method with good sensitivity and specificity for identification of such species.

REFERENCES

- [1] Shivprakash S, Radhakrishnan K, Karim PMS. *Candida* spp other than *Candida albicans*. A major cause of fungemia in a tertiary care centre. *Ind J Med Microbiol* 2007;25(4):405-407.
- [2] Mokaddas EM, Al-Sweih NA, Khan ZU. Species distribution and antifungal susceptibility of *Candida* bloodstream isolates in Kuwait: A 10-year study. *J Med Microbiol* 2007;56:255-9.
- [3] Srinivasan L, Kenneth J. Antibiotic susceptibility of *Candida* isolates in a tertiary care hospital in southern India. *Ind J Med Microbiol* 2006;24:1:80.
- [4] Golia S, Reddy KM, Karjigi KS, Hittinahalli V. Speciation of *Candida* using chromogenic and cornmeal agar with determination of fluconazole sensitivity. *Al Ameen J Med Sci* 2013;6(2):163-166.
- [5] Odds FC, Bernaerts R. CHROM agar *Candida*, a new differential isolation medium for presumptive identification of clinically important *Candida* spp. *J of Clin Microbiol* 1994;32(8):1923-9.
- [6] Jain N, Mathur P, Misra MC, Behera B, Xess I, Sharma SP. Rapid identification of yeast isolates from clinical specimens in critically ill trauma ICU patients. *J Lab Physicians* 2012;4(1):30-4.
- [7] Pinjon E, Sullivan D, Salkin I, Shanley D, Coleman D. "Simple, inexpensive, reliable method for differentiation of *Candida dubliniensis* from *Candida albicans*". *J Clin Microbiol* 1998;36(7):2093-2095.
- [8] Manjunath V, Vidya GS, Sharma A, Prakash MR, Muruges. Speciation of *Candida* by Hichrom agar and Sugar assimilation test in both HIV infected and non infected patients. *Int J Biol Med Res.* 2012;3(2):1778-1782.
- [9] Vijaya D, Harsha TR, Nagaratnamma T. *Candida* speciation using Chrom agar. *J Clin Diagn Res* 2011;5(4):755-757.
- [10] Adhikary R, Joshi S. Species distribution and anti-fungal susceptibility of *Candidaemia* at a multi super-speciality center in Southern India. *Ind J Med Microbiol* 2013;29:309-11.

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Table 1: Colour of various Candida spp. on CHROM agar for identification

	Name	Colour on Chrom agar
1.	<i>C. albicans</i>	Light green
2.	<i>C. tropicalis</i>	Metallic blue
3.	<i>C. krusei</i>	Rose pink
4.	<i>C. glabrata</i>	White

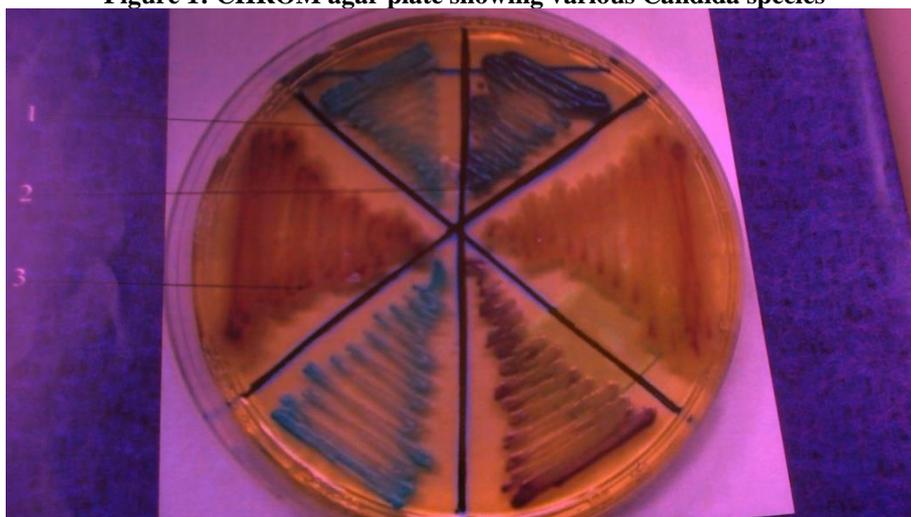
Table 2: Isolation of Candida spp. from clinical specimens

Nature of specimen	Number of <i>Candida</i> spp.	<i>Candida</i> spp. identified			
		<i>C.albicans</i>	<i>C.tropicalis</i>	<i>C.krusei</i>	<i>C.glabrata</i>
1. High vaginal swab	20	15	2	0	3
2. Urine	12	4	6	2	0
3. Stool	10	5	2	2	1
4. Blood	8	1	4	3	0
5. Sputum	3	3	0	0	0
6. Pus	3	0	1	2	0
7. Catheter tip	2	2	0	0	0
8. Ear swab	2	1	0	1	0
Total	60	31	15	10	4

Table 3: Sensitivity and specificity of CHROM agar for identification of various species of Candida

<i>Candida</i> spp.	No.of <i>Candida</i> spp. identified by conventional method	No.of <i>Candida</i> spp. identified using CHROM agar	Sensitivity of CHROM agar	Specificity of CHROM agar
1. <i>C. albicans</i>	31	32	100%	96%
2. <i>C. tropicalis</i>	15	15	100%	100%
3. <i>C. krusei</i>	10	10	100%	100%
4. <i>C. glabrata</i>	4	3	75%	100%

FIGURE

Figure 1: CHROM agar plate showing various Candida species**1. Candida albicans; 2. Candida tropicalis; 3. Candida krusei**

TABLES

Table 1: Colour of various Candida spp. on CHROM agar for identification

Name	Colour on Chrom agar
1. <i>C. albicans</i>	Light green
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		<i>C.albicans</i>	<i>C.tropicalis</i>	<i>C.krusei</i>	<i>C.glabrata</i>
9. High vaginal swab	20	15	2	0	3
10. Urine	12	4	6	2	0
11. Stool	10	5	2	2	1
12. Blood	8	1	4	3	0
13. Sputum	3	3	0	0	0
14. Pus	3	0	1	2	0
15. Catheter tip	2	2	0	0	0
16. Ear swab	2	1	0	1	0
Total	60	31	15	10	4

Table 3: Sensitivity and specificity of CHROM agar for identification of various Candida spp.

<i>Candida</i> spp.	No.of <i>Candida</i> spp. identified by conventional method	No.of spp. using agar	<i>Candida</i> identified CHROM	Sensitivity of CHROM agar	Specificity of CHROM agar
5. <i>C. albicans</i>	31	32		100%	96%
6. <i>C. tropicalis</i>	15	15		100%	100%
7. <i>C. krusei</i>	10	10		100%	100%
8. <i>C. glabrata</i>	4	3		75%	100%

Figure 1: CHROMagar plate showing various Candida species



1. *Candida albicans*
2. *Candida tropicalis*
3. *Candida krusei*